

Molecular Cloning

Associate Author

Managing Editor Editor Associate Editor Illustrator

Nina Irwin

Nancy Ford Chris Nolan Michele Ferguson Michael Ockler



Molecular Cloning A LABORATORY MANUAL

SECOND EDITION

J. Sambrook

E.F. Fritsch GENETICS INSTITUTE

T. Maniatis



Cold Spring Harbor Laboratory Press 1989

Preface to First Edition

This manual began as a collection of laboratory protocols that were used during the 1980 Cold Spring Harbor course on the Molecular Cloning of Eukaryotic Genes. These procedures had been in use in our laboratories at that time but were scattered throughout the notebooks of many different people. In 1981 we decided to produce a more complete and up-to-date manual not only for use in the next Cold Spring Harbor course, but also for eventual publication. Out of the many permutations of the methods being used, we assembled a set of "consensus protocols," which were photocopied and widely distributed to many laboratories even as the 1981 course was underway. Then in the winter of 1981–1982, the manual was substantially rewritten, and new or revised protocols and figures, as well as entirely new chapters, were added.

Even since this last rewriting, however, the field has progressed: New methods are constantly being invented and existing techniques are altered in response to changing needs. Although we have included in this manual only those protocols that have been thoroughly tested and used successfully in our laboratories, we make no claim that they are inviolable or perfect. We would welcome suggestions for improvements, and we would be grateful to be told about any new procedures that are devised.

The evolution of protocols poses the difficult problem of attribution. We have tried to give credit at appropriate places in the text to the people who originally developed the procedures presented here, but in many cases tracing a particular method to its undisputed roots has proved to be impossible. We therefore wish to apologize—and to express gratitude—to those we have been unable to acknowledge for an idea, procedure, or recipe. Our major function has been to compile, to verify, and, we hope, to clarify; less frequently we have introduced modifications, and only in rare instances have we devised new protocols. In large part, then, the manual is based on procedures developed by others, and it is to them that any credit belongs.

Because the manual was originally written to serve as a guide to those who had little experience in molecular cloning, it contains much basic material. However, the current version also deals in detail with almost every laboratory task currently used in molecular cloning. We therefore hope that newcomers to cloning and veterans alike will find material of value in this book.

Although molecular cloning seems straightforward on paper, it is more difficult to put into practice. Most protocols involve a large number of

W

individual steps and a problem with any one of them can lead the experimenter into difficulty. It is a good idea to verify the products of each step and to include controls to check the efficiency of each reaction. To deal with these problems, a well-founded understanding of the principles underlying each procedure is essential. We have therefore provided background information and references that may be useful should trouble occur.

This manual could not have been written without the help and advice of members of our laboratories and contributions from many others. We therefore wish to thank Joan Brooks, John Fiddes, Mary-Jane Gething, Tom Gingeras, David Goldberg, Steve Hughes, David Ish-Horowicz, Mike Mathews, Patty Reichel, Joe Sorge, Jim Stringer, Richard Treisman, and Nigel Whittle. We wish particularly to thank Arg Efstratiadis for his helpful discussions and criticisms of Chapter 7; Brian Seed for permission to include a description of his unpublished procedure for screening libraries by recombination (Chapter 10) and many other useful suggestions; Doug Hanahan for advice on transformation (Chapter 8); Bryan Roberts for suggestions on methods of hybridselection and cDNA cloning; Doug Melton for providing a protocol for injection of Xenopus oocytes; Ronni Greene for suggesting improvements to many protocols; Nina Irwin for providing a critical anthology of methods available for expressing eukaryotic proteins in bacteria (Chapter 12); Rich Roberts for supplying the computer analysis of the sequence of pBR322; Barbara Bachmann and Ahmad Bukhari for reviewing and correcting the list of E. coli strains; and Tom Broker, Louise Chow, Jeff Engler, and Jim Garrels for producing the elegant photographs used for the front and back covers.

We also thank all those who participated in the Cold Spring Harbor Molecular Cloning courses of 1980 and 1981. They were an excellent group of students, who struggled through the first two drafts of the manual and made many useful suggestions. We also thank Nancy Hopkins, who helped us to teach the course the first year and convinced us that producing a manual would be a worthwhile task. In 1981 Doug Engel helped teach the course and suggested many improvements to the manual. Contributing to the success of both courses were the efforts of the teaching assistants, who were Catherine O'Connell and Helen Doris Keller in the summer of 1980 and Susan Vande-Woude, Paul Bates, and Michael Weiss in 1981.

We wish to thank Patti Barkley and Marilyn Goodwin for their cheerfulness and forbearance during the typing of successive revisions of the manuscript. Our artists, Fran Cefalu and Mike Ockler, worked with great dedication and perseverance to produce the drawings for the manual. Joan Ebert kept track of the many references added to and deleted from the text and assembled the reference list. We are also grateful to Nancy Ford, Director of Publications, Cold Spring Harbor Laboratory, for her encouragement and support. Finally, without the patience, skill, and diplomacy of Doug Owen, who prepared the manuscript for the printer and helped us in many other ways, this book would not exist.

> Tom Maniatis Ed Fritsch Joe Sambrook

wi Preface to First Edition

Since the publication of the first edition of this Laboratory Manual in 1982. there has been a vast increase both in the number of people who use molecular cloning and in the range and power of the techniques used to handle recombinant DNA. This remarkable proliferation of cloning methods is reflected in the number of gene sequences in the GenBank DNA sequence data base. In 1982 there were fewer than 350 gene sequences on file, but in less than four years this number grew to almost 5000. Today over 15,000 sequences are listed. These figures, impressive as they are, do not do justice to the increased sophistication with which cloned genes are now analyzed. In 1982, respectable journals would still accept manuscripts containing little more than the partial sequence of a cDNA clone; today, the publication of a complete sequence is taken for granted, and papers describing the initial cloning of a cDNA will often also contain elegant accounts of the expression of the gene product in prokaryotic or eukaryotic hosts. In most cases, this first paper is rapidly followed by others that use site-directed mutagenesis to explore the relationship between the structure and function of the relevant protein. The cloning and analysis of elements that regulate the expression of eukaryotic genes has moved at a similar, exciting pace.

This increase in the range and speed of molecular cloning is reflected in the tripling of size of this manual and its consequent division into three volumes. Techniques that were mentioned only in passing in the first edition, such as mutagenesis, expression of cloned genes in mammalian cells, and dideoxy-mediated sequencing, are now described in depth; new sections have been added that deal with recently invented techniques, such as amplification of DNA by the polymerase chain reaction; and we have included modern variations and embellishments of many of the basic methods that were the mainstay of the first edition. We hope that any inconvenience caused by the expansion in size of the manual will be compensated by an increase in the richness of its content. We hope that this second edition will be a resource for the experienced cloner, a starting point for the student, and a guidebook for the next generation of investigators in molecular cloning.

The burgeoning of molecular cloning has also led to its commercialization, and we are now able to purchase a wide variety of high quality and reasonably priced reagents and enzymes. Although this is a very positive development, it has had a few unfortunate side effects. One is the proliferation of preassembled kits to carry out particular cloning tasks. Although these kits reduce the possibility of trivial errors, they also tend to discourage experimenters from thinking about what they are doing. It becomes all too

vii

easy to follow blindly instructions to add 2 μ l of solution A without knowing what the particular reagent is, why it is necessary, or why it is added at a particular point in the protocol. Kits therefore reward the scientifically illiterate and inhibit the development of improvements. In an attempt to counteract this trend, we have greatly increased the amount of background material at the beginning of each chapter of the manual and we have provided full references. Users of the manual who read this material should have no problem understanding either the general design or the specific details of the experimental protocols. We also strongly recommend reading the relevant protocols in their entirety before commencing work. This allows reagents to be prepared ahead of time and permits the investigator to carry out the protocol efficiently.

This manual could not have been produced without the help and encouragement of a large number of people. We are extremely grateful to colleagues from the University of Texas Southwestern Medical Center at Dallas, from Genetics Institute, from Harvard University, and from many other institutions who have read individual chapters, have submitted protocols, and have made many invaluable suggestions that have immeasurably improved the manual. We have listed all these individuals with their affiliations in a special section at the end of this preface.

We owe a special debt of thanks to Rick Myers and Alison Cowie of the University of California at San Francisco, who read all of the chapters in draft, eliminated many embarrassing errors, and made many valuable suggestions for improvements; to Winship Herr of Cold Spring Harbor Laboratory and Mary-Jane Gething of Southwestern Medical Center, who drafted the chapters on DNA sequencing and expression of cloned genes in mammalian cells, respectively; and to Judy Campbell, who contributed significantly to the preparation of the chapter on enzymes. We are also grateful to Mike Ockler, who provided all of the conceptual illustrations in this edition, and to Carolyn Doyle, who assembled the index.

Nina Irwin has played a special role in the writing of this manual. Among her many contributions are the *E. coli* strain list and the maps of the many strains of bacteriophage λ , plasmids, cosmids, and bacteriophage M13 used in this manual. These maps are based on many months of painstaking work to reconstruct and verify the genealogy of these host and vector strains. In addition, Nina also drafted the chapter on expression of cloned genes in prokaryotic hosts, read and reviewed all of the chapters in manuscript and galleys, and searched the scientific literature with great intelligence and skill to validate many of the facts that appear.

We also wish to thank Jim Watson for his continued interest in this manual. The support he has provided made this edition possible.

Finally, we would like to express our deep appreciation of the work of our editor Nancy Ford. For over two years, she has worked untiringly to bring order and sense to the entire project and to improve and clarify our writing. Nancy and her assistant Michele Ferguson have both served as a source of encouragement, sympathy, and friendship. We are greatly in their debt.

> J. Sambrook E.F. Fritsch T. Maniatis

viii Preface to Second Edition

Acknowledgments

Ed Alderman, Genetics Institute Patricia Ashley, University of Texas Southwestern Medical Center Margaret Baron, Harvard University Frank Baas, Harvard University Phil Bird, University of Texas Southwestern Medical Center Tom Bittick, University of Texas Southwestern Medical Center Colleen Brewer, University of Texas Southwestern Medical Center Gene Brown, Genetics Institute H. Franklin Bunn, Harvard Medical School Steve Clark, Genetics Institute Preston Dunnmon, University of Texas Southwestern Medical Center Anne Ephrussi, Harvard University Henry Erlich, Perkin Elmer Cetus Chen-Ming Fan, Harvard University Ken Ferguson, Cold Spring Harbor Laboratory Bill Garrard, University of Texas Southwestern Medical Center Bob Gerard, University of Texas Southwestern Medical Center Gary Gilliland, Harvard Medical School Doug Hanahan, University of California, San Francisco Ed Harlow, Cold Spring Harbor Laboratory Jean Henneberry, University of Texas Southwestern Medical Center Bill Huse, Stratagene David Ish-Horowicz, Imperial Cancer Research Fund David Israel, Genetics Institute Randy Kaufman, Genetics Institute Andrew Keller, Harvard University John Knopf, Genetics Institute Steve Lacey, University of Texas Southwestern Medical Center Ray MacDonald, University of Texas Southwestern Medical Center Ed Madison, University of Texas Southwestern Medical Center Mike Mathews, Cold Spring Harbor Laboratory Michael McClelland, University of Chicago John McCoy, Genetics Institute Steve McKnight, Carnegie Institution of Washington Doug Melton, Harvard University Alan Michelson, Harvard University George Morris, Genetics Institute Michael Nelson, University of Chicago Robin Reed, Harvard University Rich Roberts, Cold Spring Harbor Laboratory Laura Roman, University of Texas Southwestern Medical Center Susan Rosenberg, University of Utah David Russell, University of Texas Southwestern Medical Center Brian Seed, Harvard Medical School Conrad Seghers, University of Texas Southwestern Medical Center Mark Segal, University of Texas Southwestern Medical Center Chuck Shoemaker, Harvard School of Public Health Huda Shubeita, University of Texas Southwestern Medical Center Harinder Singh, University of Chicago Beth Smith, Information Center, Genetics Institute Joe Sorge, Stratagene Lisa Sultzman, Genetics Institute Galvin Swift, University of Texas Southwestern Medical Center Patty Temple, Genetics Institute Jeff Vieira, Rutgers University Geoff Wahl, Salk Institute Steve Wasserman, University of Texas Southwestern Medical Center Gordon Wong, Genetics Institute Rick Young, Massachusetts Institute of Technology Jian-Hua Zhang, University of Texas Southwestern Medical Center Mark Zoller, Genentech

ix

Contents

BOOK 1

Plasmid Vectors

Essential Features of Plasmids 1.3

Replication and Incompatibility 1.3 Mobilization 1.5 Selectable Markers 1.5

Plasmid Vectors 1.7

P

DEVELOPMENT OF PLASMID CLONING VECTORS 1.7

Plasmid Vectors That Permit Histochemical Identification of Recombinant Clones 1.8

Plasmid Vectors Carrying Origins of Replication Derived from Single-stranded Bacteriophages 1.9

Plasmid Vectors Carrying Bacteriophage Promoters 1.9

Plasmid Vectors That Allow Direct Selection of Recombinant Clones 1.9 Plasmid Expression Vectors 1.10

COMMONLY USED PLASMID VECTORS 1.11

pBR322 1.12

pUC18, pUC19 1.13 pUC118, pUC19 1.14 pSP64, pSP65, pGEM-3, pGEM-3Z, pGEM-3Zf(-), pGEM-4, pGEM-4Z 1.15 π AN13 1.19 BLUESCRIPT M13 + , M13 - 1.20

Extraction and Purification of Plasmid DNA 1.21

Growth of the Bacterial Culture 1.21 Harvesting and Lysis of the Bacteria 1.22 Purification of Plasmid DNA 1.23

SMALL-SCALE PREPARATIONS OF PLASMID DNA 1.25 Harvesting and Lysis of Bacteria 1.25

xi

HARVESTING 1.25 LYSIS BY ALKALI 1.25 LYSIS BY BOILING 1.29

Solving Problems That Arise with Minipreparations of Plasmid DNA 1.31 Rapid Disruption of Bacterial Colonies to Test the Size of Plasmids 1.32

LARGE-SCALE PREPARATIONS OF PLASMID DNA 1.33

Amplification of Plasmids in Rich Medium 1.33 Harvesting and Lysis of Bacteria 1.34

HARVESTING 1.34 LYSIS BY BOILING 1.34 LYSIS BY SODIUM DODECYL SULFATE 1.36 LYSIS BY ALKALI 1.38

PURIFICATION OF PLASMID DNA 1.40

Purification of Plasmid DNA by Precipitation with Polyethylene Glycol 1.40 Purification of Closed Circular DNA by Equilibrium Centrifugation in CsCl-Ethidium Bromide Gradients 1.42 CONTINUOUS GRADIENTS 1.42 DISCONTINUOUS GRADIENTS 1.44

Removal of Ethidium Bromide from DNAs Purified by Equilibrium Centrifugation in CsCl-Ethidium Bromide Gradients 1.46 METHOD 1: EXTRACTION WITH ORGANIC SOLVENTS 1.46 METHOD 2: ION-EXCHANGE CHROMATOGRAPHY 1.47

Decontamination of Ethidium Bromide Solutions 1.49 DECONTAMINATION OF CONCENTRATED SOLUTIONS OF ETHIDIUM BROMIDE 1.49 DECONTAMINATION OF DILUTE SOLUTIONS OF ETHIDIUM BROMIDE 1.50

Removal of RNA from Preparations of Plasmid DNA 1.51 CENTRIFUGATION THROUGH 1 M NaCl 1.51 CHROMATOGRAPHY THROUGH BIO-GEL A-150m OR SEPHAROSE CL-4B 1.52

Strategies for Cloning in Plasmid Vectors 1.53

STRATEGIES FOR LIGATION 1.53

The Nature of the Ends of the Foreign DNA Fragment 1.53 FRAGMENTS CARRYING NONCOMPLEMENTARY PROTRUDING TERMINI 1.53 FRAGMENTS CARRYING IDENTICAL TERMINI (BLUNT-ENDED OR PROTRUDING) 1.56 FRAGMENTS CARRYING BLUNT ENDS 1.56

The Nature of the Restriction Sites in the Plasmid Vector and the Foreign DNA 1.59

DEPHOSPHORYLATION OF LINEARIZED PLASMID DNA 1.60 Test Ligations and Transformations 1.62

LIGATION REACTIONS 1.63

Setting Up Ligation Reactions Between Foreign DNA and Plasmid Vectors 1.63 Ligation of Cohesive Termini 1.68 Ligation of Blunt-ended DNA 1.70 CONDENSING AGENTS 1.70 Rapid Cloning in Plasmid Vectors 1.72

Preparation and Transformation of Competent E. coli 1.74

Transformation of E. coli by High-voltage Electroporation (Electrotransformation) 1.75 Protocol I: Preparation of Fresh or Frozen Competent E. coli 1.76 Protocol II: Fresh Competent E. coli Prepared Using Calcium Chloride 1.82

xii Contents

Identification of Bacterial Colonies That Contain Recombinant Plasmids 1.85

RESTRICTION ANALYSIS OF SMALL-SCALE PREPARATIONS OF PLASMID DNA 1.85

α-COMPLEMENTATION 1.85 Testing Bacteria for α-Complementation 1.86

INSERTIONAL INACTIVATION 1.87

SCREENING BY HYBRIDIZATION 1.90

Transferring Small Numbers of Colonies to Nitrocellulose Filters 1.92 Replicating Colonies onto Nitrocellulose Filters 1.93

METHOD 1 1.93

METHOD 2 1.96

Lysis of Colonies and Binding of DNA to Nitrocellulose Filters 1.98 METHOD 1 1.98

METHOD 2 1.100

Hybridization to Nitrocellulose Filters Containing Replicas of Bacterial Colonies 1.101

References 1.105

2

Bacteriophage λ Vectors

Molecular Biology of Bacteriophage λ 2.3

THE LYTIC CYCLE 2.3 Adsorption 2.3 Immediate Early Transcription 2.5 Delayed Early Transcription 2.5 DNA Replication 2.5 Late Transcription 2.5 Assembly 2.7 Lysis 2.8

LYSOGENY 2.8

Bacteriophage λ Vectors 2.9

CONSTRUCTION OF BACTERIOPHAGE λ VECTORS: A BRIEF HISTORY 2.9 CHOOSING THE APPROPRIATE BACTERIOPHAGE λ VECTOR 2.11

MAPS OF BACTERIOPHAGE λ VECTORS 2.15

 λ^+ 2.17 CHARON 4A 2.18 CHARON 21A 2.20 CHARON 32 2.22 CHARON 33 2.24

Contents xiii

CHARON 34 2.26 CHARON 35 2.28 CHARON 40 2.30 EMBL3A 2.32 $\lambda 2001$ 2.34 $\lambda DASH$ 2.36 λFIX 2.38 $\lambda gt10$ 2.40 $\lambda gt11$ 2.42 $\lambda gt18$ 2.44 $\lambda gt20$ 2.46 $\lambda gt22$ 2.48 $\lambda ORF8$ 2.50 $\lambda ZAP/R$ 2.52

CHOOSING A BACTERIAL HOST FOR BACTERIOPHAGE λ VECTORS 2.55 Restriction and Modification 2.55 Amber Suppressors 2.55 Recombination Systems 2.55

Bacteriophage λ Growth, Purification, and DNA Extraction 2.60

PLAQUE PURIFICATION OF BACTERIOPHAGE λ 2.60 Preparation of Plating Bacteria 2.60 Plating Bacteriophage λ 2.61 Picking Bacteriophage λ Plaques 2.63

PREPARING STOCKS OF BACTERIOPHAGE λ FROM SINGLE PLAQUES 2.64 Plate Lysate Stocks 2.64 PREPARATION OF PLATE LYSATE STOCKS: PROTOCOL I 2.65 PREPARATION OF PLATE LYSATE STOCKS: PROTOCOL II 2.66 Small-scale Liquid Cultures 2.67 Long-term Storage of Bacteriophage λ Stocks 2.68

LARGE-SCALE PREPARATION OF BACTERIOPHAGE λ 2.69 Infection at Low Multiplicity 2.70 Infection at High Multiplicity 2.72

PURIFICATION OF BACTERIOPHAGE λ 2.73Standard Method for Purification of Bacteriophage λ 2.73Alternative Methods for Purification of Bacteriophage λ 2.77PELLETING BACTERIOPHAGE PARTICLES2.77GLYCEROL STEP GRADIENT2.78EQUILIBRIUM CENTRIFUGATION IN CESIUM CHLORIDE2.79

EXTRACTION OF BACTERIOPHAGE & DNA 2.80

Cloning in Bacteriophage λ 2.82

PREPARATION OF VECTOR DNA 2.82
 Digestion of Bacteriophage λ DNA with Restriction Enzymes 2.83
 Purification of Bacteriophage λ Arms 2.85
 CENTRIFUGATION THROUGH SUCROSE DENSITY GRADIENTS 2.85
 CENTRIFUGATION THROUGH SODIUM CHLORIDE GRADIENTS 2.88
 Preparation of Vectors Treated with Alkaline Phosphatase 2.90
 Digestion of Bacteriophage λ Vectors with Two Restriction Enzymes 2.92
 Ligation of Bacteriophage λ Arms to Fragments of Foreign DNA 2.94

PACKAGING OF BACTERIOPHAGE & DNA IN VITRO 2.95

Maintenance and Testing of Lysogens of Bacteriophage λ 2.96

Preparation of Packaging Extracts and Packaging of Bacteriophage λ DNA In Vitro 2.98

PROTOCOL I: PREPARATION OF PACKAGING EXTRACTS FROM TWO LYSOGENS 2.100 PROTOCOL I: PACKAGING IN VITRO USING TWO EXTRACTS 2.104 PROTOCOL II: PREPARATION OF PACKAGING EXTRACTS FROM ONE LYSOGEN 2.105 PROTOCOL II: PACKAGING IN VITRO USING ONE EXTRACT 2.107

Identification and Analysis of Recombinants 2.108

IN SITU HYBRIDIZATION OF BACTERIOPHAGE & PLAQUES 2.108

Immobilization of Bacteriophage λ Plaques on Nitrocellulose Filters or Nylon Membranes 2.109

Immobilization of Bacteriophage λ Plaques on Nitrocellulose Filters Following In Situ Amplification 2.112

Hybridization to Nitrocellulose Filters Containing Replicas of Bacteriophage λ Plaques 2.114

RAPID ANALYSIS OF BACTERIOPHAGE *λ* ISOLATES 2.118 Plate Lysate Method 2.118 Liquid Culture Method 2.121

References 2.122

3

Cosmid Vectors

Cloning in Cosmid Vectors 3.5

Cosmid Vectors 3.7

COSMID VECTORS FOR PROPAGATION OF EUKARYOTIC DNA IN BACTERIA 3.9

pJB8 3.9 c2RB 3.13 pcos1EMBL 3.17

COSMID VECTORS FOR TRANSFECTION OF MAMMALIAN CELLS 3.18 pHC79-2cos/tk 3.19 pCV103, pCV107, pCV108 3.19 pTM, pMCS, pNNL 3.19 pHSG274 3.19

cos202, cos203 3.19 pWE15, pWE16 3.21 CHAROMID 9 VECTORS 3.25

Construction of Genomic DNA Libraries in Cosmid Vectors 3.27 CLONING IN PHOSPHATASE-TREATED COSMID VECTORS 3.28

Contents XV

Preparation of Vector DNA 3.28 TEST LIGATION 3.29 Partial Digestion of Eukaryotic DNA with MboI or Sau3AI 3.32 Ligation and Packaging 3.33

CLONING IN COSMID VECTORS DIGESTED WITH TWO RESTRICTION ENZYMES AND TREATED WITH PHOSPHATASE 3.35 Preparation of Vector DNA 3.36 Treatment of Eukaryotic DNA with Alkaline Phosphatase 3.38 Ligation and Packaging 3.40

PREPARATION OF VECTORS CONTAINING TWO cos SITES 3.42

Amplification and Storage of Cosmid Libraries 3.44

Replica Filters 3.46 Amplification of Cosmid Libraries in Liquid Culture 3.50 Preparation of a Transducing Lysate of Packaged Cosmids 3.52

Problems Commonly Encountered When Using Cosmids 3.54

References 3.56

Single-stranded, Filamentous Bacteriophage Vectors

The Biology of Filamentous Bacteriophages 4.3

Filamentous Bacteriophages as Vectors 4.7

BACTERIOPHAGE M13 VECTORS 4.7

BACTERIAL HOSTS FOR BACTERIOPHAGE M13 VECTORS 4.12

PROBLEMS COMMONLY ENCOUNTERED WHEN CLONING IN FILAMENTOUS BACTERIOPHAGE VECTORS 4.16

PHAGEMIDS: PLASMIDS CONTAINING AN ORIGIN OF REPLICATION DERIVED FROM A FILAMENTOUS BACTERIOPHAGE 4.17

THE USES OF SINGLE-STRANDED DNA GENERATED BY RECOMBINANTS CONSTRUCTED IN PHAGEMID AND FILAMENTOUS BACTERIOPHAGE VECTORS 4.20

Propagation of Bacteriophage M13 and Preparation of DNA 4.21

PLAQUE PURIFICATION OF BACTERIOPHAGE M13 4.21 Preparation of Plating Bacteria 4.21

xvi Contents

Plating Bacteriophage M13 4.22 Picking Bacteriophage M13 Plaques 4.24

PREPARING STOCKS OF BACTERIOPHAGE M13 FROM SINGLE PLAQUES 4.25 Liquid Cultures 4.25

PREPARATION OF BACTERIOPHAGE M13 DNA 4.26

Small-scale Preparations of the Replicative Form of Bacteriophage M13 DNA 4.26

Small-scale Preparations of Single-stranded Bacteriophage M13 DNA 4.29 Large-scale Preparations of the Replicative Form of Bacteriophage M13 DNA 4.31

Large-scale Preparations of Single-stranded Bacteriophage M13 DNA 4.32

Cloning into Bacteriophage M13 Vectors and Transfection of Competent Bacteria 4.33

Cloning DNA Fragments into Bacteriophage M13 Vectors 4.35 Preparation of Competent Bacteria 4.36 Transfection of Competent Bacteria with Bacteriophage M13 DNA 4.37

Identification and Analysis of Recombinants 4.39

Direct Gel Electrophoresis 4.39 In Situ Hybridization of Bacteriophage M13 Plaques 4.41 Analysis of Small-scale Preparations of the Replicative Form of Bacteriophage M13 DNA 4.41 Testing the Orientation of DNA Cloned in Bacteriophage M13 Vectors 4.42

Cloning in Phagemids 4.44

Growth of M13K07 4.47 Production of Single-stranded Phagemid DNA 4.48 Screening Colonies by Superinfection 4.49

References 4.51

5

Enzymes Used in Molecular Cloning

Restriction and DNA Methylation Enzymes 5.3

LIGATION OF TERMINI CREATED BY RESTRICTION ENZYMES 5.10 Compatible Cohesive Termini 5.10 Blunt Ends 5.11 Incompatible Cohesive Termini 5.11

ISOSCHIZOMERS 5.14

Contents xvii

DNA METHYLATION 5.15

Methylation by Commonly Used Strains of E. coli 5.15 dam METHYLASE 5.15 dcm METHYLASE 5.15 METHYLATION-DEPENDENT RESTRICTION SYSTEMS IN E. coli 5.15

M.EcoRI METHYLASE 5.16 Modification of Restriction Sites by DNA Methylation 5.16 Influence of Methylation on DNA Mapping 5.26

DIGESTING DNA WITH RESTRICTION ENZYMES 5.28 Setting Up Digestions with Restriction Enzymes 5.31

Other Enzymes Used in Molecular Cloning 5.33

DNA POLYMERASES 5.35

DNA Polymerase I (Holoenzyme) 5.36 Large Fragment of DNA Polymerase I (Klenow Fragment) 5.40 Bacteriophage T4 DNA Polymerase 5.44 Bacteriophage T7 DNA Polymerase 5.48 Modified Bacteriophage T7 DNA Polymerase (SequenaseTM) 5.49 Taq DNA Polymerase and AmpliTaqTM 5.50 Reverse Transcriptase (RNA-dependent DNA Polymerase) 5.52 Terminal Transferase (Terminal Deoxynucleotidyl Transferase) 5.56

DNA-DEPENDENT RNA POLYMERASES 5.58 Bacteriophage SP6 and Bacteriophages T7 and T3 RNA Polymerases 5.58

LIGASES, KINASES, AND PHOSPHATASES 5.61 Bacteriophage T4 DNA Ligase 5.62 E. coli DNA Ligase 5.64 Bacteriophage T4 RNA Ligase 5.66 Bacteriophage T4 Polynucleotide Kinase 5.68 Alkaline Phosphatases 5.72

NUCLEASES 5.73 Nuclease BAL 31 5.73 Nuclease S1 5.78 Mung-bean Nuclease 5.80 Ribonuclease A 5.81 Ribonuclease T1 5.82 Deoxyribonuclease I 5.83 Exonuclease III 5.84 Bacteriophage λ Exonuclease 5.86

DNA-binding Proteins 5.87

Single-stranded DNA-binding Protein (SSB) 5.87 RecA Protein 5.88 Topoisomerase I 5.89

References 5.90

Gel Electrophoresis of DNA

ĥ

Agarose Gel Electrophoresis 6.3

Factors Affecting the Rate of DNA Migration in Agarose Gels 6.4 MOLECULAR SIZE OF THE DNA 6.4 AGAROSE CONCENTRATION 6.4 CONFORMATION OF THE DNA 6.5 APPLIED VOLTAGE 6.6 DIRECTION OF THE ELECTRIC FIELD 6.6 BASE COMPOSITION AND TEMPERATURE 6.6 PRESENCE OF INTERCALATING DYES 6.6 COMPOSITION OF THE ELECTROPHORESIS BUFFER 6.6 Apparatuses Used for Agarose Gel Electrophoresis 6.8

PREPARATION AND EXAMINATION OF AGAROSE GELS 6.9 Preparation of an Agarose Gel 6.9

Minigels 6.14 Staining DNA in Agarose Gels 6.15

Decontamination of Ethidium Bromide Solutions 6.16 DECONTAMINATION OF CONCENTRATED SOLUTIONS OF ETHIDIUM BROMIDE 6.16 DECONTAMINATION OF DILUTE SOLUTIONS OF ETHIDIUM BROMIDE 6.17 Photography 6.19 Alkaline Agarose Gels 6.20

RECOVERY AND PURIFICATION OF DNA FRACTIONATED ON AGAROSE GELS 6.22

Electrophoresis onto DEAE-cellulose Membrane 6.24 Electroelution into Dialysis Bags 6.28 Recovery of DNA from Low-melting-temperature Agarose Gels 6.30 Purification of DNA Recovered from Agarose Gels 6.32 PASSAGE THROUGH DEAE-SEPHACEL 6.32 EXTRACTION WITH ORGANIC SOLVENTS 6.34

Polyacrylamide Gel Electrophoresis 6.36

PREPARATION OF NONDENATURING POLYACRYLAMIDE GELS 6.39

DETECTION OF DNA IN POLYACRYLAMIDE GELS 6.44 Staining with Ethidium Bromide 6.44 Autoradiography 6.45 UNFIXED, WET GELS 6.45 FIXED, DRIED GELS 6.45

ISOLATION OF DNA FRAGMENTS FROM POLYACRYLAMIDE GELS 6.46 "Crush and Soak" Method 6.46

Other Types of Gels 6.49

STRAND SEPARATING GELS 6.49

DENATURING GRADIENT POLYACRYLAMIDE GELS 6.49

Contents xix

PULSED-FIELD GEL ELECTROPHORESIS6.50Design of the Apparatus6.51Staining DNA Separated by Pulsed-field Gel Electrophoresis6.52Preparation of DNA for Pulsed-field Gel Electrophoresis6.53ISOLATION OF INTACT DNA FROM MAMMALIAN CELLS6.53ISOLATION OF INTACT DNA FROM YEAST6.55RESTRICTION ENZYME DIGESTION OF DNA IN AGAROSE BLOCKS6.57Markers for Pulsed-field Gel Electrophoresis6.58

References 6.60

Extraction, Purification, and Analysis of Messenger RNA from Eukaryotic Cells

Extraction and Purification of RNA 7.3

CONTROLLING RIBONUCLEASE ACTIVITY 7.3

Laboratory Procedures 7.3 Inhibitors of Ribonucleases 7.4 Methods That Disrupt Cells and Inactivate Ribonucleases Simultaneously 7.5

ISOLATION OF RNAs 7.6

Isolation of Total RNA from Mammalian Cells 7.6 Rapid Isolation of Total RNA from Mammalian Cells 7.10 Isolation of Cytoplasmic RNA from Mammalian Cells 7.12 Isolation of Total RNA from Eggs and Embryos 7.16 Isolation of Total RNA by Extraction with Strong Denaturants 7.18 EXTRACTION OF RNA WITH GUANIDINIUM THIOCYANATE FOLLOWED BY CENTRIFUGATION IN CESIUM CHLORIDE SOLUTIONS 7.19 EXTRACTION OF RNA WITH GUANIDINE HCI AND ORGANIC SOLVENTS 7.23

SELECTION OF POLY(A)⁺ RNA 7.26

FRACTIONATION OF RNA BY SIZE IN THE PRESENCE OF METHYLMERCURIC HYDROXIDE 7.30

Electrophoresis of RNA through Agarose Gels Containing Methylmercuric Hydroxide 7.31

RECOVERY OF RNA FROM AGAROSE GELS CONTAINING METHYLMERCURIC HYDROXIDE 7.33

Fractionation of RNA by Centrifugation through Sucrose Gradients Containing Methylmercuric Hydroxide 7.35

Analysis of RNA 7.37

NORTHERN HYBRIDIZATION 7.39

Electrophoresis of RNA after Denaturation with Glyoxal and Dimethyl Sulfoxide 7.40

Electrophoresis of RNA through Gels Containing Formaldehyde 7.43 Transfer of Denatured RNA to Nitrocellulose Filters 7.46 Transfer of Denatured RNA to Nylon Membranes 7.49 Staininging RNA Before and After Transfer to Nitrocellulose Filters 7.51 METHOD 1 7.51 METHOD 2 7.51

Hybridization and Autoradiography 7.52

DOT AND SLOT HYBRIDIZATION OF RNA 7.53 Slot Hybridization of RNA 7.54 Slot Hybridization of Cytoplasmic RNA 7.56

MAPPING RNA WITH NUCLEASE S1 7.58 Mapping of RNA with Nuclease S1 and Double-stranded DNA Probes 7.62 Mapping of RNA with Nuclease S1 and Single-stranded DNA Probes 7.66

MAPPING OF RNA WITH RIBONUCLEASE AND RADIOLABELED RNA PROBES 7.71

ANALYSIS OF RNA BY PRIMER EXTENSION 7.79

References 7.84

Index

Contents xxii

8

Construction and Analysis of cDNA Libraries

Strategies for cDNA Cloning 8.3

PREPARATION OF mRNA FOR cDNA CLONING 8.3 Source of the mRNA 8.3 Integrity of the mRNA 8.4 Abundant mRNAs 8.6 Low-abundance mRNAs 8.6 Methods of Enrichment 8.6 FRACTIONATION OF mRNA BY SIZE 8.8 FRACTIONATION OF cDNA 8.8 IMMUNOLOGICAL PURIFICATION OF POLYSOMES 8.9

SYNTHESIS OF THE FIRST STRAND OF cDNA 8.11

SYNTHESIS OF THE SECOND STRAND OF cDNA 8.14 Self-priming 8.14 Replacement Synthesis of the Second Strand of cDNA 8.15 Primed Synthesis of the Second Strand of cDNA 8.16

MOLECULAR CLONING OF DOUBLE-STRANDED cDNA 8.21

Homopolymeric Tailing 8.21 Synthetic DNA Linkers and Adapters 8.23 Alternative Methods of Cloning cDNA 8.27 mRNA-cDNA CLONING 8.27 SEQUENTIAL ADDITION OF DIFFERENT LINKERS 8.27 cDNA CLONING IN OKAYAMA-BERG VECTORS 8.29 cDNA CLONING WITH PRIMER-ADAPTERS 8.30 cDNA CLONING IN SINGLE-STRANDED VECTORS 8.32

BACTERIOPHAGE & VECTORS USED FOR CLONING OF cDNA 8.36

Bacteriophage λ Vectors $\lambda gt10$ and $\lambda gt11$ 8.36

 $\lambda gt10 8.36 \\ \lambda gt11 8.37$

Other Bacteriophage λ Vectors Used for cDNA Cloning 8.39 λ ORF8 8.39

 λgt18, λgt19
 8.41

 λgt20, λgt21
 8.42

 λgt22, λgt23
 8.42

 λZAP
 8.44

IDENTIFICATION OF cDNA CLONES OF INTEREST 8.46 Methods of Screening 8.46 NUCLEIC ACID HYBRIDIZATION 8.46 IMMUNOLOGICAL DETECTION OF SPECIFIC ANTIGENS 8.49 SIB SELECTION OF cDNA CLONES 8.50 Methods to Validate Clones of cDNA 8.51

Protocols for cDNA Cloning 8.53

CONSTRUCTING cDNA LIBRARIES IN BACTERIOPHAGE λ VECTORS 8.54 Precautions 8.54 Protocol for the Synthesis of the First Strand of cDNA 8.60 Protocol for the Synthesis of the Second Strand of cDNA 8.64 Methylation of cDNA 8.66 Ligation to Synthetic Phosphorylated Linkers 8.68 Size Selection of cDNA 8.70 Ligation to Bacteriophage λ Arms 8.73 Analysis of cDNA Inserts 8.76 Generation of a Complete cDNA Library 8.77 Amplification of cDNA Libraries 8.78 AMPLIFICATION OF LIBRARIES CONSTRUCTED IN BACTERIOPHAGE λgt10: SELECTION AGAINST PARENTAL BACTERIOPHAGES 8.78 AMPLIFICATION OF LIBRARIES CONSTRUCTED IN BACTERIOPHAGE λgt11 AND ITS DERIVATIVES AND λZAP OR λZAPII 8.79

PROBLEMS COMMONLY ENCOUNTERED WITH cDNA CLONING 8.80

References 8.82

9

Analysis and Cloning of Eukaryotic Genomic DNA

Vectors Used to Construct Eukaryotic Genomic DNA Libraries 9.4

Bacteriophage λ and Cosmid Vectors 9.4 The Yeast Artificial Chromosome System 9.5

BACTERIOPHAGE λ VECTORS RECOMMENDED FOR CONSTRUCTION OF LIBRARIES OF EUKARYOTIC GENOMIC DNA 9.7

COSMID VECTORS RECOMMENDED FOR CONSTRUCTION OF LIBRARIES OF EUKARYOTIC GENOMIC DNA 9.13

Isolation of High-molecular-weight DNA from Mammalian Cells 9.14

Isolation of DNA from Mammalian Cells: Protocol I 9.16 Isolation of DNA from Mammalian Cells: Protocol II 9.20 Isolation of DNA from Mammalian Cells: Protocol III 9.22

Partial Digestion of High-molecular-weight Eukaryotic DNA with Restriction Enzymes 9.24

Pilot Experiments 9.24 Large-scale Preparation of Partially Digested DNA 9.27 Partial Filling of Recessed 3' Termini of Fragments of Genomic DNA 9.29 Amplification of a Genomic DNA Library 9.30

Contents xxiii

Analysis of Genomic DNA by Southern Hybridization 9.31

SEPARATION OF RESTRICTION FRAGMENTS OF MAMMALIAN GENOMIC DNA BY AGAROSE GEL ELECTROPHORESIS 9.32

TRANSFER OF DNA FROM AGAROSE GELS TO SOLID SUPPORTS 9.34 Transfer of DNA to Nitrocellulose Filters 9.38

- CAPILLARY TRANSFER OF DNA TO NITROCELLULOSE FILTERS 9.38 SIMULTANEOUS TRANSFER OF DNA FROM A SINGLE AGAROSE GEL TO TWO NITROCELLULOSE FILTERS 9.41
- Transfer of DNA from Agarose Gels to Nylon Membranes 9.42 CAPILLARY TRANSFER OF DNA TO NYLON MEMBRANES UNDER NEUTRAL CONDITIONS 9.44

CAPILLARY TRANSFER OF DNA TO NYLON MEMBRANES UNDER ALKALINE CONDITIONS $\ 9.45$

HYBRIDIZATION OF RADIOLABELED PROBES TO IMMOBILIZED NUCLEIC ACIDS 9.47

Hybridization of Radiolabeled Probes to Nucleic Acids Immobilized on Nitrocellulose Filters or Nylon Membranes 9.52

Hybridization of Radiolabeled Oligonucleotides to Genomic DNA 9.56 Removal of Radiolabeled Probes from Nitrocellulose Filters and Nylon Membranes 9.58

REMOVING PROBES FROM NITROCELLULOSE FILTERS 9.58 REMOVING PROBES FROM NYLON MEMBRANES 9.58

References 9.59

10

Preparation of Radiolabeled DNA and RNA Probes

Synthesis of Uniformly Labeled Double-stranded DNA Probes 10.6

NICK TRANSLATION OF DNA 10.6 Stock Solutions Used in Nick Translation 10.7 Protocol for Nick Translation 10.8 Alternative Protocol for Nick Translation 10.11

SYNTHESIS OF UNIFORMLY LABELED DNA PROBES USING RANDOM OLIGONUCLEOTIDE PRIMERS 10.13 Synthesis of Probes from Denatured Double-stranded DNA 10.14 SYNTHESIS OF RADIOLABELED PROBES BY PRIMER EXTENSION 10.14 RADIOLABELING OF DNA IN THE PRESENCE OF MELTED AGAROSE 10.16

Preparation of Single-stranded Probes 10.18

PRODUCTION OF SINGLE-STRANDED DNA PROBES USING BACTERIOPHAGE M13 VECTORS 10.19
Primer:Template Ratios and Nucleotide Concentrations 10.20
Synthesis of Single-stranded DNA Probes 10.22
Isolation of Small (<150 Nucleotides) Probes by Alkaline Chromatography on Sepharose CL-4B 10.25
Troubleshooting 10.26

xxiv Contents

SYNTHESIS OF RNA PROBES BY IN VITRO TRANSCRIPTION OF DOUBLE-STRANDED DNA TEMPLATES BY BACTERIOPHAGE DNA-DEPENDENT RNA POLYMERASES 10.27 Plasmid Vectors for Preparing RNA Probes 10.29 Preparation of DNA Template 10.31 Synthesis of RNA In Vitro 10.32

Synthesis of RNA Probes Radiolabeled to High Specific Activity 10.34 Troubleshooting 10.36

SYNTHESIS OF cDNA PROBES 10.38

Identification of cDNA Clones 10.38

DIFFERENTIAL SCREENING 10.38

ABSORBED PROBES 10.40

SUBTRACTED LIBRARIES 10.40

SUBTRACTED PROBES RADIOLABELED TO HIGH SPECIFIC ACTIVITY 10.43

Synthesis of Total cDNA Probes Complementary to Single-stranded RNA Using Oligonucleotides as Primers 10.44

Synthesis of Radiolabeled, Subtracted cDNA Probes Using Oligo(dT) as a Primer 10.46

Synthesis of Subtracted Probes Radiolabeled to High Specific Activity 10.48

Labeling the 5' and 3' Termini of DNA 10.51

LABELING THE 3' TERMINI OF DOUBLE-STRANDED DNA USING THE KLENOW FRAGMENT OF E. coli DNA POLYMERASE I 10.51

LABELING THE 3' TERMINI OF DOUBLE-STRANDED DNA WITH BACTERIOPHAGE T4 DNA POLYMERASE 10.54 Rapid End-labeling of DNA 10.54 Replacement Synthesis 10.56

LABELING THE 5' TERMINUS OF DNA WITH BACTERIOPHAGE T4 POLYNUCLEOTIDE KINASE 10.59

Forward Reaction 10.60

USING DNA MOLECULES WITH PROTRUDING 5' TERMINI AS SUBSTRATES 10.60 USING DNA MOLECULES WITH BLUNT ENDS OR RECESSED 5' TERMINI AS SUBSTRATES 10.62

DEPHOSPHORYLATION OF DNA 10.64

Exchange Reaction 10.66

USING DNA MOLECULES WITH PROTRUDING 5'-PHOSPHATE TERMINI AS TEMPLATES 10.66

References 10.68

Synthetic Oligonucleotide Probes

Types and Uses of Oligonucleotide Probes 11.3

SINGLE OLIGONUCLEOTIDES OF DEFINED SEQUENCE 11.4

POOLS OF SHORT OLIGONUCLEOTIDES WHOSE SEQUENCES ARE HIGHLY DEGENERATE 11.5

Contents XXV

The Effects of Length and Degeneracy of the Oligonucleotide on the Specificity of Hybridization 11.7

Designing Degenerate Pools of Short Oligonucleotides 11.9

POOLS OF LONGER OLIGONUCLEOTIDES OF LESSER DEGENERACY 11.11 Guessmers 11.11 DESIGNING A GUESSMER 11.11 LABELING OF GUESSMERS 11.15 Oligonucleotides That Contain a Neutral Base at Positions of Degeneracy 11.17

Purification and Radiolabeling of Synthetic Oligonucleotides 11.20

PURIFICATION OF SYNTHETIC OLIGONUCLEOTIDES 11.21

Preparation of Synthetic Oligonucleotides 11.21

Recovery of Synthetic Oligonucleotides by Electrophoresis Through a Denaturing Polyacrylamide Gel 11.23

Isolation of Oligonucleotides by Reversed-phase Chromatography on a Silica Gel 11.29

LABELING OF SYNTHETIC OLIGONUCLEOTIDES BY PHOSPHORYLATION WITH BACTERIOPHAGE T4 POLYNUCLEOTIDE KINASE 11.31

PURIFICATION OF RADIOLABELED SYNTHETIC

OLIGONUCLEOTIDES 11.33

Purification of Radiolabeled Oligonucleotides by Precipitation with Ethanol 11.34

- Purification of Radiolabeled Oligonucleotides by Precipitation with Cetylpyridinium Bromide 11.35
- Purification of Radiolabeled Oligonucleotides by Chromatography Through Bio-Gel P-60 11.37

Purification of Radiolabeled Oligonucleotides by Chromatography on a Sep-Pak C₁₈ Column 11.39

LABELING OF SYNTHETIC OLIGONUCLEOTIDES USING THE KLENOW FRAGMENT OF E. coli DNA POLYMERASE I 11.40

Conditions for Hybridization of Oligonucleotide Probes 11.45

CALCULATING MELTING TEMPERATURES FOR PERFECTLY MATCHED HYBRIDS BETWEEN OLIGONUCLEOTIDES AND THEIR TARGET SEQUENCES 11.46

ESTIMATING THE EFFECTS OF MISMATCHES 11.47

HYBRIDIZATION OF POOLS OF OLIGONUCLEOTIDES 11.48

Preparation and Use of Solvents Containing Quaternary Alkylammonium Salts 11.50

HYBRIDIZATION OF GUESSMERS 11.52

HYBRIDIZATION OF OLIGONUCLEOTIDES THAT CONTAIN A NEUTRAL BASE AT POSITIONS OF DEGENERACY 11.54

EMPIRICAL DETERMINATION OF MELTING TEMPERATURE 11.55

References 11.58

xxvi Contents

Screening Expression Libraries with Antibodies and Oligonucleotides

Constructing Expression Libraries in Plasmid and Bacteriophage λ Vectors 12.4

 The Relative Advantages of Plasmid and Bacteriophage λ Expression Vectors 12.4
 Genomic DNA and cDNA Expression Libraries 12.6
 Expression Libraries Constructed in Plasmids 12.8
 Expression Libraries Constructed in Bacteriophage λ 12.10

Using Antibodies in Immunological Screening 12.11

Choosing the Antibody 12.11 Purification of Antisera 12.13 Methods Used to Detect Antibodies Bound to Proteins Expressed in E. coli 12.14 Validation of Clones Isolated by Immunological Screening 12.15

Immunological Screening of Expression Libraries 12.16

SCREENING EXPRESSION LIBRARIES CONSTRUCTED IN BACTERIOPHAGE λ VECTORS 12.16

SCREENING BACTERIAL COLONIES 12.21

Preparation of Colonies for Screening 12.21 METHOD 1 12.21

METHOD 2 12.23

Processing Filters for Immunological Screening of Colonies 12.24

REMOVING ANTI-E. coli ANTIBODIES BY PSEUDOSCREENING 12.25

PREPARATION OF E. coli LYSATES FOR ABSORPTION OF ANTI-E. coli ANTIBODIES 12.26

REMOVAL OF ANTI-E. coli ANTIBODIES BY AFFINITY CHROMATOGRAPHY 12.27

RADIOIODINATION OF IMMUNOGLOBULIN G 12.29

Screening cDNA Libraries Constructed in Bacteriophage λ Expression Vectors with Synthetic Oligonucleotides 12.30

Preparing Radiolabeled Concatenated Probes 12.32
Preparation of Filters for Screening with Radiolabeled Concatenated Probes 12.34
Probing Immobilized Proteins with Radiolabeled DNA 12.36
Preparation of Lysates Containing Fusion Proteins Encoded by Bacteriophage λgt11 Lysogens 12.38

References 12.41

Contents xxvii

13

DNA Sequencing

Sequencing Techniques and Strategies 13.3

SANGER METHOD OF DIDEOXY-MEDIATED CHAIN TERMINATION 13.6 Reagents Used in the Sanger Method of DNA Sequencing 13.6 PRIMERS 13.6 TEMPLATES 13.7 DNA POLYMERASES 13.7 RADIOLABELED dNTPs 13.9 ANALOGS OF dNTPs 13.10

MAXAM-GILBERT CHEMICAL DEGRADATION OF DNA METHOD 13.11

SEQUENCING STRATEGIES 13.14 Confirmatory Sequencing 13.14 De Novo Sequencing 13.14 FACTORS AFFECTING THE CHOICE BETWEEN RANDOM AND DIRECTED STRATEGIES 13.18

Random Sequencing 13.21

GENERATION OF A LIBRARY OF RANDOMLY OVERLAPPING CLONES 13.24
Purification and Ligation of the Target DNA 13.24
Fragmentation of the Target DNA 13.26
SONICATION 13.26
DIGESTION WITH DNAase I IN THE PRESENCE OF MANGANESE IONS 13.28
Repair and Size Selection of DNA 13.30
Preparation of Vector DNA 13.31
Ligation to Vector DNA 13.33

Directed Sequencing 13.34

GENERATION OF NESTED SETS OF DELETION MUTANTS 13.34 Generation of Nested Sets of Deletions with Exonuclease III 13.39

Sequencing by the Sanger Dideoxy-mediated Chain-termination Method 13.42

SETTING UP DIDEOXY-MEDIATED SEQUENCING REACTIONS 13.42 Preparation of Single-stranded DNA 13.42 Preparation of Primers 13.42 Microtiter Plates 13.42 Chain-extension/Chain-termination Reaction Mixtures 13.43 STOCK SOLUTIONS OF dNTPs AND ddNTPs 13.44

DENATURING POLYACRYLAMIDE GELS 13.45 Preparation of Buffer-gradient Polyacrylamide Gels 13.47 Loading and Running Gradient Sequencing Gels 13.54 Autoradiography of Sequencing Gels 13.56 Reading the Sequence 13.58

xxviii Contents

DIDEOXY-MEDIATED SEQUENCING REACTIONS USING THE KLENOW FRAGMENT OF E. coli DNA POLYMERASE I 13.59 Preparation 13.59 PREPARATION OF WORKING SOLUTIONS OF dNTPs 13.60

PREPARATION OF WORKING SOLUTIONS OF dNTPs 13.60 PREPARATION OF WORKING SOLUTIONS OF ddNTPs 13.60 Sequencing Reactions 13.61

DIDEOXY-MEDIATED SEQUENCING REACTIONS USING SEQUENASES 13.65 Preparation 13.65 Sequencing Reactions 13.67

SEQUENCING DENATURED DOUBLE-STRANDED DNA TEMPLATES 13.70 Sequencing of Plasmid DNAs Purified by Equilibrium Centrifugation in CsCl– Ethidium Bromide Gradients 13.71

Removal of RNA from Minipreparations of Plasmid DNA by Precipitation with Lithium Chloride 13.72

PROBLEMS THAT ARISE WITH DIDEOXY-MEDIATED SEQUENCING 13.73 Template-specific Problems 13.73 Systematic Problems 13.73 Problems with Polyacrylamide Gels 13.74

Sequencing by the Maxam-Gilbert Method 13.78

Asymmetric Labeling of Target DNA 13.78 Preparation of Target DNA for Maxam-Gilbert Sequencing 13.83 Reagents, Solutions, and Apparatuses 13.83

THE TRADITIONAL METHOD OF MAXAM-GILBERT SEQUENCING 13.88 Cleavage at G Residues 13.88 Cleavage at Purine Residues (A + G) 13.90 Cleavage at Pyrimidine Residues (C + T) 13.91 Cleavage at C Residues 13.92 Cleavage at A and C Residues (A > C) 13.93 Treatment of Samples with Piperidine 13.94

ALTERNATIVE METHODS OF MAXAM-GILBERT SEQUENCING 13.95

TROUBLESHOOTING GUIDE FOR MAXAM-GILBERT SEQUENCING 13.95 Reading Sequencing Gels 13.95 Problems Commonly Encountered 13.95

References 13.102

14

In Vitro Amplification of DNA by the Polymerase Chain Reaction

APPLICATIONS OF PCR AMPLIFICATION 14.5 Generation of Specific Sequences of Cloned Double-stranded DNA for Use as Probes 14.6

Contents xxix

Generation of Probes Specific for Uncloned Genes by Selective Amplification of Particular Segments of cDNA 14.7
Generation of Libraries of cDNA from Small Amounts of mRNA 14.9
Generation of Large Amounts of DNA for Sequencing 14.10
Analysis of Mutations 14.11
Chromosome Crawling 14.12

AMPLIFICATION METHODS 14.14

Precautions 14.14

Components of the Polymerase Chain Reaction 14.15 OLIGONUCLEOTIDES 14.15 BUFFERS USED FOR POLYMERASE CHAIN REACTIONS 14.15

Taq DNA POLYMERASE 14.16 DEOXYRIBONUCLEOSIDE TRIPHOSPHATES 14.16

TARGET SEQUENCES 14.16

Amplification Reactions 14.18

Amplification of DNA Generated by Reverse Transcription of mRNA 14.20

SEQUENCING AMPLIFIED DNA BY THE SANGER DIDEOXY-MEDIATED CHAIN-TERMINATION METHOD 14.22

Protocol I: Sequencing Amplified DNA with Radiolabeled Oligonucleotide Primers 14.22

REMOVAL OF OLIGONUCLEOTIDES AND EXCESS dNTPs FROM AMPLIFIED DNA BY SPIN DIALYSIS 14.22

RADIOLABELING OF THE OLIGONUCLEOTIDE SEQUENCING PRIMER $\ 14.25$ ANNEALING $\ 14.26$

Protocol II: Sequencing Single-stranded DNA Templates Generated by Asymmetric Amplification 14.28

QUANTITATION OF INITIAL CONCENTRATION OF TARGET SEQUENCES 14.30

References 14.34



Site-directed Mutagenesis of Cloned DNA

Generation of Deletions and Insertions 15.3

SIMPLE DELETIONS OR INSERTIONS 15.3

SYSTEMATIC DELETIONS AND INSERTIONS 15.5

Linker-insertion Mutagenesis 15.5

GENERATION OF LINKER-INSERTION MUTANTS WITH FREQUENTLY CUTTING RESTRICTION ENZYMES 15.8

Generation of Nested Sets of Deletion Mutants 15.14

GENERATION OF BIDIRECTIONAL SETS OF DELETION MUTANTS BY DIGESTION WITH NUCLEASE BAL 31 15.20

CLEAVAGE OF DOUBLE-STRANDED CLOSED CIRCULAR DNA WITH PANCREATIC DNAase I IN THE PRESENCE OF $\rm Mn^{++}~15.27$

Linker-scanning Mutagenesis 15.32

ISOLATION OF TARGET FRAGMENTS CONTAINING A BglII LINKER AND A kan^r GENE 15.37

RECOVERY OF THE TARGET DNA FRAGMENT 15.47

EXCISION OF THE TARGET DNA 15.48

ANALYSIS OF CLONES THAT CONTAIN A BglII LINKER IN THE TARGET REGION 15.50

XXX Contents

Oligonucleotide-mediated Mutagenesis 15.51

Preparation of Single-stranded Target DNA 15.53 Design and Selection of Mutagenic Oligonucleotides 15.54 Hybridization of Oligonucleotides to the Template DNA and Primer Extension 15.57 Transfection of E. coli and Screening for Mutants 15.59 Recovery of the Mutated Fragment of DNA 15.60 Methods to Improve the Efficiency of Oligonucleotide-mediated

Mutagenesis 15.61

OLIGONUCLEOTIDE-MEDIATED MUTAGENESIS BY THE DOUBLE-PRIMER METHOD 15.63

SCREENING BACTERIOPHAGE M13 PLAQUES AND BACTERIAL COLONIES BY HYBRIDIZATION TO RADIOLABELED OLIGONUCLEOTIDES 15.66 Radiolabeling of Oligonucleotides by Phosphorylation 15.66 Screening Bacteriophage M13 Plaques by Hybridization to Radiolabeled Oligonucleotides 15.68

Screening Bacterial Colonies by Hybridization to Radiolabeled Oligonucleotides 15.72

OLIGONUCLEOTIDE-MEDIATED MUTAGENESIS BY SELECTION AGAINST TEMPLATE STRANDS THAT CONTAIN URACIL (KUNKEL METHOD) 15.74 Preparation of Uracil-containing Single-stranded DNA 15.76

SOLVING PROBLEMS THAT ARISE IN OLIGONUCLEOTIDE-MEDIATED, SITE-DIRECTED MUTAGENESIS 15.80

Using Mutagenesis to Study Proteins 15.81

INSERTION OF HEXAMERIC LINKERS INTO PROTEIN-CODING SEQUENCES 15.85 Designing the Hexameric Linker 15.86 Insertion of Hexameric Linkers 15.88

CREATING MANY MUTATIONS IN A DEFINED SEGMENT OF DNA 15.95 Use of Degenerate Pools of Mutagenic Oligonucleotides 15.96 Treatment of Double-stranded DNA with Chemical Mutagens 15.105 Treatment of Single-stranded DNA with Sodium Bisulfite 15.106 Treatment of Single-stranded DNA with Chemicals That Damage All Four Bases 15.107 Misincorporation of Nucleotides by DNA Polymerases 15.108

References 15.109

Index

Contents XXXXI

BOOK 3

16

Expression of Cloned Genes in Cultured Mammalian Cells

Expression of Proteins 16.3

EXPRESSION OF PROTEINS FROM CLONED GENES 16.3

FUNCTIONAL COMPONENTS OF MAMMALIAN EXPRESSION VECTORS 16.5

Prokaryotic Plasmid Sequences That Facilitate the Construction, Propagation, and Amplification of Recombinant Vector Sequences in Bacteria 16.5

A Eukaryotic Expression Module That Contains All of the Elements Required for the Expression of Foreign DNA Sequences in Eukaryotic Cells 16.5 PROMOTER AND ENHANCER ELEMENTS 16.5 TERMINATION AND POLYADENYLATION SIGNALS 16.6 SPLICING SIGNALS 16.7

ELEMENTS FOR REPLICATION AND SELECTION 16.8

Foreign DNA Sequences 16.15

VECTOR SYSTEMS 16.17

Plasmid-based Vectors That Do Not Carry a Eukaryotic Replicon 16.17
Plasmid DNA Expression Vectors Containing Regulatory Elements from Eukaryotic Viruses 16.17
SIMIAN VIRUS 40 VECTORS 16.17
BOVINE PAPILLOMAVIRUS VECTORS 16.23
EPSTEIN-BARR VIRUS VECTORS 16.26
Amplification Systems 16.28

Introduction of Recombinant Vectors into Mammalian Cells 16.30

TRANSFECTION OF COPRECIPITATES OF CALCIUM PHOSPHATE AND DNA 16.32

Standard Protocol for Calcium Phosphate-mediated Transfection of Adherent Cells 16.33

Calcium Phosphate-mediated Transfection of Adherent Cells in Suspension 16.37

Calcium Phosphate-mediated Transfection of Cells Growing in Suspension 16.38 Modified Calcium Phosphate-mediated Transfection Procedure 16.39

TRANSFECTION MEDIATED BY DEAE-DEXTRAN 16.41 Transfection Using DEAE-Dextran: Protocol I 16.42 Transfection Using DEAE-Dextran: Protocol II 16.45

DNA TRANSFECTION USING POLYBRENE 16.47

DNA TRANSFECTION BY PROTOPLAST FUSION 16.48 Preparation of Protoplasts 16.49 Fusion of Protoplasts to Adherent Mammalian Cells 16.50 Fusion of Protoplasts to Mammalian Cells Growing in Suspension 16.52

DNA TRANSFECTION BY ELECTROPORATION 16.54

xxxii Contents

Strategies for Studying Gene Regulation 16.56

VECTORS CARRYING REPORTER GENES 16.57

ASSAYS FOR CHLORAMPHENICOL ACETYLTRANSFERASE AND β-GALACTOSIDASE ACTIVITIES 16.59 Preparation of Extracts 16.59

Assays for Chloramphenicol Acetyltransferase 16.60 METHOD 1: THIN-LAYER CHROMATOGRAPHY 16.60

METHOD 2: EXTRACTION WITH ORGANIC SOLVENTS 16.63

METHOD 3: DIFFUSION OF REACTION PRODUCTS INTO SCINTILLATION FLUID 16.64 Assay for β -Galactosidase in Extracts of Mammalian Cells 16.66

Cloning by Expression in Mammalian Cells 16.68

Cloning by Expression of cDNA Clones 16.69 Expression Cloning of Genomic DNA 16.70

References 16.73

17 Expression of Cloned Genes in Escherichia coli

Production of Fusion Proteins 17.3

Vector Systems for the Expression of lacZ Fusion Genes 17.4 Construction of Expression Plasmids and Detection of Fusion Proteins 17.8 Preparation of Fusion Proteins for Antibody Production 17.9

Production of Intact Native Proteins 17.10

EXPRESSION OF PROKARYOTIC GENES: PROMOTERS 17.11 The Bacteriophage λ p_L Promoter 17.11 The trp-lac Promoter 17.13 The Bacteriophage T7 Promoter 17.15

EXPRESSION OF EUKARYOTIC GENES: PROMOTERS AND RIBOSOME-BINDING SITES 17.17 Preparation of a DNA Fragment for Placement Adjacent to a Functional Ribosome-binding Site 17.18 SYNTHESIS OF DNA ENCODING THE AMINO TERMINUS 17.18 PRIMER REPAIR 17.18 ENGINEERING A RESTRICTION SITE 17.22 Expression of a Gene from the Prepared Fragments 17.25

ALTERNATIVE EXPRESSION SYSTEMS 17.29 Expression of a Cloned Gene as Part of a Fusion Protein That Can Be Cleaved by a Protease or Cyanogen Bromide 17.29 PRODUCTION OF HYBRID PROTEINS THAT CAN BE CLEAVED WITH FACTOR X_a 17.30 Expression of Secreted Foreign Proteins 17.31 phoA-MEDIATED EXPRESSION AND SECRETION 17.32

Contents xxxiii

QUANTITATING THE LEVELS OF EXPRESSION OF CLONED GENES 17.34 Monitoring Expression by β-Galactosidase Activity 17.35

INCREASING EXPRESSION OF CLONED GENES 17.36

PROTEIN PURIFICATION 17.37

Inclusion Bodies 17.37

CELL LYSIS 17.38

PURIFICATION AND WASHING OF INCLUSION BODIES: METHOD 1 17.39 PURIFICATION AND WASHING OF INCLUSION BODIES: METHOD 2 17.40 SOLUBILIZATION OF INCLUSION BODIES 17.41

References 17.42

18

Detection and Analysis of Proteins Expressed from Cloned Genes

Production of Antibodies 18.3

Factors Affecting Immune Response 18.3 SPECIES OF ANIMAL 18.3 GENETIC FACTORS 18.4 PHYSICAL STATE OF THE ANTIGEN 18.4 AMOUNT OF ANTIGEN 18.4 ROUTE OF INJECTION 18.5 IMMUNIZATION SCHEDULES 18.5 Immunizing with Small Amounts of Antigen 18.6 Monoclonal Antibodies 18.7 Raising Antisera Against Synthetic Peptides 18.7 COUPLING OF SYNTHETIC PEPTIDES TO KEYHOLE LIMPET HEMOCYANIN 18.8 Collection and Storage of Antisera 18.10

Purification of Antibodies 18.11

PURIFICATION OF ANTIBODIES BY ADSORPTION TO PROTEIN A 18.12

PURIFICATION OF ANTIBODIES BY ABSORPTION 18.14 Removal of Cross-reacting Antibodies from Antisera 18.15 Purification of Immunospecific Antibodies 18.16 AFFINITY PURIFICATION OF MONOSPECIFIC ANTIBODIES USING ANTIGEN IMMOBILIZED ON NITROCELLULOSE FILTERS 18.17

Immunological Assays 18.19

SOLID-PHASE RADIOIMMUNOASSAY 18.19 Solid-phase Radioimmunoassays Using Two Antibodies 18.21 Iodination of Antibodies 18.24 RADIOIODINATION OF ANTIBODIES USING THE CHLORAMINE-T METHOD 18.24

IMMUNOPRECIPITATION 18.26

xxxiv Contents

Radiolabeling the Target Protein 18.26

RADIOLABELING MAMMALIAN CELLS WITH [³⁵S]METHIONINE AND [³⁵S]CYSTEINE 18.27

METABOLIC RADIOLABELING OF PROTEINS EXPRESSED IN YEASTS AND BACTERIA 18.29

Lysis of Cells 18.30

LYSIS OF CULTURED MAMMALIAN CELLS 18.34

MECHANICAL LYSIS OF YEAST 18.35

ENZYMATIC LYSIS OF YEAST 18.36 RAPID LYSIS OF YEAST CELLS 18.38

LYSIS OF BACTERIA 18.40

Formation of Antigen-Antibody Complexes 18.42 PRECLEARING THE CELL LYSATE 18.43

Immunoprecipitation of the Target Protein 18.44

SDS-Polyacrylamide Gel Electrophoresis of Proteins 18.47

PREPARATION OF SDS-POLYACRYLAMIDE GELS 18.49

POURING SDS-POLYACRYLAMIDE GELS 18.51

STAINING SDS-POLYACRYLAMIDE GELS WITH COOMASSIE BRILLIANT BLUE 18.55

STAINING SDS-POLYACRYLAMIDE GELS WITH SILVER SALTS 18.56 DRYING SDS-POLYACRYLAMIDE GELS 18.58

TRANSFER OF PROTEINS FROM SDS-POLYACRYLAMIDE GELS TO SOLID SUPPORTS: IMMUNOLOGICAL DETECTION OF IMMOBILIZED PROTEINS (WESTERN BLOTTING) 18.60

Preparation and Electrophoresis of Samples 18.61

LYSIS OF MAMMALIAN CELLS AND TISSUE IN GEL-LOADING BUFFER 18.62 Transfer of Proteins from SDS-Polyacrylamide Gels to Solid Supports 18.64

Staining Proteins Immobilized on Nitrocellulose Filters 18.67

STAINING WITH PONCEAU S 18.67

STAINING WITH INDIA INK 18.68

Blocking Binding Sites for Immunoglobulins on the Nitrocellulose Filter 18.69 Binding of the Primary Antibody to the Target Protein 18.70

INCUBATING THE NITROCELLULOSE FILTER WITH THE PRIMARY ANTIBODY DIRECTED AGAINST THE TARGET PROTEIN 18.70

INCUBATING THE NITROCELLULOSE FILTER WITH THE SECONDARY IMMUNOLOGICAL REAGENT 18.72

USE OF CHROMOGENIC SUBSTRATES WITH ENZYME-COUPLED ANTIBODIES 18.74

Translation of mRNAs 18.76

TRANSLATION OF RNA IN RETICULOCYTE LYSATES 18.76 Preparation of Rabbit Reticulocyte Lysate 18.77 Translation of Reticulocyte Lysates 18.79

TRANSLATION OF SYNTHETIC mRNAs IN VITRO 18.81 Synthesis of Synthetic mRNAs 18.82 Translation of Synthetic RNAs 18.85

References 18.86

Contents XXXXV

A ppendixes

Appendix A: Bacterial Media, Antibiotics, and Bacterial Strains

LIQUID MEDIA A.1

LB Medium (Luria-Bertani Medium) A.1 NZCYM Medium A.1 NZYM Medium A.1 NZM Medium A.1 Terrific Broth A.2 SOB Medium A.2 SOC Medium A.2 2×YT Medium A.3 M9 Minimal Medium A.3

MEDIA CONTAINING AGAR OR AGAROSE A.4

STORAGE MEDIA A.5 Stab Cultures A.5 Cultures Containing Glycerol A.5 BACTERIAL CULTURES GROWING IN LIQUID MEDIA A.5 BACTERIAL CULTURES GROWING ON AGAR PLATES A.5

ANTIBIOTICS A.6

SOLUTIONS FOR WORKING WITH BACTERIOPHAGE λ A.7 Maltose A.7 SM A.7 TM A.7 λ Diluent A.8

BACTERIAL STRAIN LIST A.9

Appendix B: Preparation of Reagents and Buffers Used in Molecular Cloning

CONCENTRATIONS OF ACIDS AND BASES B.1 pK_as of Commonly Used Buffers B.1 Preparation of Tris Buffers of Various Desired pH Values B.1 Concentrations of Acids and Bases: Common Commercial Strengths B.2 Approximate pH Values for Various Concentrations of Stock Solutions B.3

PREPARATION OF ORGANIC REAGENTS B.4 Phenol B.4 EQUILIBRATION OF PHENOL B.4 Phenol:Chloroform:Isoamyl Alcohol (25:24:1) B.5

ATOMIC WEIGHTS AND ISOTOPIC DATA B.6

STOCK SOLUTIONS AND BLOCKING AGENTS B.9

ENZYMES B.16 Proteolytic Enzymes B.16 Lysozyme B.17

XXXVI Contents

RNAase That Is Free of DNAase B.17 DNAase That Is Free of RNAase B.17 AFFINITY CHROMATOGRAPHY ON AGAROSE 5'-(4-AMINOPHENYLPHOSPHORYL) URIDINE-2'(3')-PHOSPHATE B.17 ADSORPTION TO MACALOID B.18 HEATING IN THE PRESENCE OF IODOACETATE B.19

COMMONLY USED BUFFERS B.20 Phosphate Buffers B.21 Alkaline Lysis Buffers for Minipreparations of Plasmid DNA B.22 Electrophoresis Buffers B.23 Enzyme Buffers B.26

Appendix C: Properties of Nucleic Acids

VITAL STATISTICS OF DNA C.1 Haploid DNA Content of Various Organisms C.1 Concentration of DNA in Solution C.1 B Form of DNA C.2 Relationship Between the Length of DNA and Its Molecular Weight C.3

PURINES AND PYRIMIDINES C.3

Numbering of Atoms C.3 Adenine and Related Compounds C.4 Cytosine and Related Compounds C.6 Guanine and Related Compounds C.8 Thymine and Related Compounds C.10 Uracil and Related Compounds C.11 Unusual Bases C.12 Nucleoside Analogs Used as Chain Terminators in DNA Sequencing C.13 Nucleotide Sequence Data Banks C.14

Appendix D: Codons and Amino Acids

The Genetic Code (Nuclear Genes) D.1 Prokaryotic Supressors of Nonsense Mutations Used in Molecular Cloning D.1 Properties of Amino Acids D.2 Classification of Amino Acids D.6

Appendix E: Commonly Used Techniques in Molecular Cloning

GLASSWARE AND PLASTICWARE E.1 Siliconizing Glassware, Plasticware, and Glass Wool E.1

PURIFICATION OF NUCLEIC ACIDS E.3 Extraction with Phenol:Chloroform E.3

QUANTITATION OF DNA AND RNA E.5

Spectrophotometric Determination of the Amount of DNA or RNA E.5 Ethidium Bromide Fluorescent Quantitation of the Amount of Double-stranded DNA E.5 SARAN WRAP METHOD E.6

AGAROSE PLATE METHOD E.6 MINIGEL METHOD E.6

DECONTAMINATION OF ETHIDIUM BROMIDE SOLUTIONS E.8

Contents xxxvii

Decontamination of Concentrated Solutions of Ethidium Bromide (i.e., solutions containing > 0.5 mg/ml) E.8

METHOD 1 E.8 METHOD 2 E.8

Decontamination of Dilute Solutions of Ethidium Bromide (e.g., electrophoresis buffer containing 0.5 µg/ml ethidium bromide) E.9 METHOD 1 E.9 METHOD 2 E.9

CONCENTRATING NUCLEIC ACIDS E.10

Precipitation with Ethanol or Isopropanol E.10 PRECIPITATION OF DNA IN MICROFUGE TUBES E.12 PRECIPITATION OF RNA WITH ETHANOL E.15 PRECIPITATION OF LARGE RNAS WITH LITHIUM CHLORIDE E.15 CONCENTRATING NUCLEIC ACIDS BY EXTRACTION WITH BUTANOL E.16

DRYING DOWN ³²P-LABELED NUCLEOTIDES FROM MIXTURES OF ETHANOL AND WATER E.17

MEASUREMENT OF RADIOACTIVITY IN NUCLEIC ACIDS E.18 Precipitation of Nucleic Acids with Trichloroacetic Acid E.18 Adsorption to DE-81 Filters E.19

STANDARD MARKERS FOR GEL ELECTROPHORESIS E.20

AUTORADIOGRAPHY E.21 Fluorography E.24 Sensitivity of Different Autoradiographic Methods E.25 Setting up Autoradiographs E.26

SEPARATION OF SINGLE-STRANDED AND DOUBLE-STRANDED DNA BY HYDROXYAPATITE CHROMATOGRAPHY E.30

GEL-FILTRATION CHROMATOGRAPHY E.34 Preparation of Sephadex E.35 Column Chromatography E.36

SPUN-COLUMN CHROMATOGRAPHY E.37

PREPARATION OF DIALYSIS TUBING E.39

Appendix F: Subcloning

FILLING RECESSED 3' TERMINI F.2

REMOVING PROTRUDING 3' TERMINI F.4

RAPID CLONING IN PLASMID VECTORS F.6

ADDITION OF LINKERS TO BLUNT-ENDED DNA F.8 Enzymatic Phosphorylation of Nonphosphorylated Linkers F.8 Ligation of Phosphorylated Linkers to Blunt-ended Target Fragments F.9

Appendix G: List of Suppliers

References

Index

XXXVIII Contents