

## Oxazaphosphorine Cytostatics: Past–Present–Future

Seventh Cain Memorial Award Lecture<sup>1</sup>

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## Abstract

The development of the oxazaphosphorine cytostatics cyclophosphamide, ifosfamide, and trofosfamide was based on the idea of applying the transport form/active form principle to the highly reactive nitrogen mustard group. A critical analysis and synopsis of the available results and knowledge will include examination of the extent to which the hypotheses on which this concept is based have been confirmed by experimental and clinical findings:

1. Chemical synthesis succeeded in converting the reactive nitrogen mustard into an inactive transport form (latentionation).

2. The requirement that the transport form be enzymatically activated to the active form in the target organ (the cancer cell) has been achieved by a sequence of metabolic reactions.

3. The aim of considerably increasing the therapeutic index of alkylating agents has been achieved by the oxazaphosphorine cytostatics. The greater cancerotoxic selectivity is closely correlated with the cytotoxic specificity of their activated primary metabolites.

4. The cancerotoxic selectivity of oxazaphosphorines was further increased when mesna was introduced as a regional uroprotector. Mesna eliminates the risk of therapy-limiting urotoxic side effects of oxazaphosphorines. With mesna protection, these cytostatics can be given in higher doses with increased safety, and their therapeutic efficacy can be enhanced.

5. Stabilization of the primary oxazaphosphorines, e.g., by attaching 2-mercaptoethanesulfonic acid (mafosfamide), opens up new possibilities in preclinical investigations and in therapy, e.g., for the clonogenic stem cell assay, for *in vitro* purging in autologous bone marrow transplantation, for regional perfusion of tumors, and, in small doses, for immunomodulation, where appropriate, in conjunction with "biological response modifiers."

## Introduction

The great distinction conferred by the Cain Memorial Award is evident from its stated objective "to give recognition for outstanding preclinical research leading to the discovery of a significant new therapeutic agent for the improved care of cancer patients." It is thus a great pleasure for me to join the list of award winners, whose contributions are regarded as milestones in the advance of cancer chemotherapy.

The acknowledgement of the pharmacologist's work in the development of the oxazaphosphorine cytostatics must take into account a number of scientists from other specialties who, in the past, have made essential contributions, and it suggests that the future prospects in this area of research should be outlined: "To see the future requires the memory of the past".<sup>2</sup>

## Environment and Personal Background

Among the personalities I remember with particular respect from the 1930's when I was becoming a physician and researcher are the physician Ernst Edens from Düsseldorf and the pharmacologists Wolfgang Heubner and Hermann Druckrey in Ber-

lin. In the words of Heubner, "I love that word conscientiousness. It conveys the indispensable association between scientific knowledge and responsibility—*scientia et conscientia*". This was one of the essential principles of Wolfgang Heubner at the Pharmacology Institute of the University of Berlin, and I have attempted to base my life and work on this too. Hermann Druckrey was responsible for my broad introduction to the fundamentals of pharmacology, and specifically to cancer research. He was one of the founders of the field of oncological pharmacology, the scientific basis of which he developed and considerably extended. Even in these early years in Berlin we were interested in drugs to inhibit cell division, and a particularly suitable subject for the systematic study of this was the fertilized sea urchin egg, being simple and not subject to pain. The results of these studies at the Zoological Research Station in Naples with the support of the greatly respected Reinhard Dohrn proved subsequently to be a useful basis for the chemotherapy research at ASTA.

I became head of the Pharmacology Department of Asta-Werke in 1949. This allowed me to place my specialist's knowledge of pharmacology and internal medicine at the service of versatile drug research and drug development. From the outset our aim was not to leave the discovery of new drugs to chance but to base it on rational scientific hypotheses and to take account of the results of biophysical and biochemical, as well as pharmacological and clinical, research. Together with Hermann Druckrey in Freiburg and Berthold Schneider in Hannover, we developed the pharmacological and biometric bases for the screening and pharmacological characterization of new drugs. This included standardization of the experimental conditions for determination of chemical and biological activities *in vitro*. Optimization of the pharmacological testing *in vivo* was based on the creation of adequate test models for determining the therapeutic and toxic effects, on the formulation of sharper action criteria, on the use of quantitative test methods, especially of the dose-response analysis, and on the development of pharmacodynamic and pharmacokinetic investigation methods for the determination of the cumulative properties. The evaluation was based on the determination of the therapeutic index, of the danger coefficient, and of the therapeutic units (1).

Despite this good scientific basis, it was a very bold step for Asta-Werke, a medium-sized pharmaceutical company in Bielefeld, to have, in 1952, the chemist Herbert Arnold, the pharmacologist Norbert Brock, and the clinician Hilmar Wilmanns, together with Ewald Kipper, the founder and then president of the company, decide to make the chemotherapy of malignant diseases a main aim of research. The prospects for the development of cancer chemotherapy were regarded in the early 1950's as rather poor, and the benefit/risk relation was generally assessed as extremely unfavorable. To achieve a breakthrough it was necessary in this area of drug therapy, more than in any other, to conduct long-term and high-level research and to provide sufficient finance, and the latter probably appeared at that time scarcely possible for a company of the size of Asta-Werke. Indeed most research into cancer chemotherapy was then, worldwide, confined almost entirely to the large cancer research centers,

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<sup>2</sup> W. Dürching, personal communication.

which received governmental support; only a few pharmaceutical companies were carrying out their own cancer research. Asta-Werke has kept faith with its decision for more than 35 years now, the only pharmaceutical company in Germany to do so.

To gain this objective it was necessary to have a group of workers able to see and to some extent master the problems in this area. ASTA then had had competent chemical, pharmacological, and clinical research departments. For experimental oncology, immunology, biochemistry, and biometry we sought cooperations with universities who provided continuous conceptual and methodological support. Thus, our research group can be regarded as an early and very efficiently functioning model of targeted collaboration between researchers in industry and the universities, and I would like to pay a grateful tribute to the pharmacologist and oncologist Hermann Druckrey of Freiburg, the biochemist Hans-Jürgen Hohorst of Frankfurt, the microbiologist and immunologist Jürgen Potel of Bielefeld and Hannover, and the biometrician Berthold Schneider of Hannover for many years of friendly cooperation.

### Transport Form/Active Form as a Therapeutic Principle

We were aware from the outset that, in view of the financial limits, it was not possible to carry out an extensive screening program analogous to the testing of thousands of substances each year by the large national cancer research centers. The important point for our research group was to go beyond the empirical procedure which had predominated hitherto and to base the development of cancer chemotherapy on theoretical concepts (2) or, as Wolfgang Heubner had said, to follow "creative ideas" (3). Our aim in this was to develop substances with a greater selectivity of cancerotoxic action and to reduce the risks to patients by increasing the therapeutic range. This became the guiding principle of our subsequent work.

Druckrey proposed that a reactive, toxic drug should be administered not in the active form but in a chemically masked, inactive transport form. This should be converted into the active form in the body where possible, preferentially in the tumor cell (4). The first successful implementation of this idea was fosfestrol (Honvan), the diphosphate derivative of diethylstilbestrol, which is still widely used in the treatment of metastatic prostate carcinoma.

From the outset we regarded the basic idea as a hypothesis or theory, the validity of which can be examined objectively (2). The history of the oxazaphosphorine cytostatics is a good example of the interplay between idea and reality since intensive research worldwide in the decades following the formulation of the primary idea has confirmed the concept by experimental and clinical studies and by specific investigations. Three ideas are crucially involved:

1. The extension of the "transport form/active form" principle to highly reactive nitrogen mustards; this resulted in the oxazaphosphorine cytostatics cyclophosphamide, ifosfamide, and trofosfamide.

2. The introduction of the terms "cytotoxic specificity" and "cancerotoxic selectivity" exemplified by the oxazaphosphorines. This made the mechanism of action of this compound class understandable and established their special position in the group of alkylating agents.

3. The development of regional, and thus of an organ-specific, detoxification of the urotoxic oxazaphosphorine cytostatics by detailed investigation of the metabolism and pharmacokinetics; this resulted in a further considerable increase in the cancerotoxic selectivity.

### Oxazaphosphorine Cytostatics

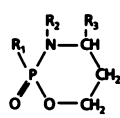
After the success of fosfestrol, our team, including Arnold, Bourseaux, and Wilmanns, tried to apply the transport form/active form principle to the group of alkylating nitrogen mustards. The structure of these compounds appeared to be particularly suitable for designing a prodrug (5). Their cytotoxic action is closely related to the reactivity of the 2-chloroethyl groups, which in turn is linked with the basicity of the central nitrogen atom. Thus, chemical synthesis can be used to alter the reactivity of the functional groups in a specific manner. Ishidate *et al.* (6) were the first to demonstrate this relationship by describing chlormethine *N*-oxide as an antitumor drug.

One of the earliest attempts to utilize chemically derived structural variations of the nitrogen mustard molecule for the synthesis of compounds with greater tumor selectivity was the work of Friedman and Seligman with open-chain phosphoramidate mustard compounds (7). These compounds are chemically stable but, according to our own experiments (8), unsuitable as substrates for enzymatic activation; their oncocidal efficacy was insufficient, resulting in a narrow therapeutic range.

**Cyclophosphamide.** Our approach was aimed at obtaining cyclic *N*-phosphorylation products of *nor*-nitrogen mustard as prodrugs. These chemically and pharmacologically inactive transport forms should offer opportunities for enzymatic activation to the cancerotoxic active form, *e.g.*, by the incorporation of phosphoramidate and phosphoric ester bonds into the molecule.

These novel compounds were synthesized by reacting *N,N*-bis(2-chloroethyl)phosphoramidate dichloride with  $\alpha,\omega$ -alkanolamines of various chain lengths. Further cyclic variants were produced with appropriate bifunctional alkanes yielding mono-, di-, or triamides of phosphoric acid (9, 10). From the pharmacotherapeutic viewpoint, the diamides, especially the nitrogen mustard phosphoramidate esters (or oxazaphosphorines), were most attractive since they were largely inactive chemically and biologically *in vitro* but highly active *in vivo*. Of the more than 1000 derivatives that were synthesized, some compounds were found to possess particularly favorable properties (Fig. 1). Cyclophosphamide, the prototype of this class, was the first oxazaphosphorine cytostatic which accorded with theory; it was a chemically and pharmacologically inactive transport form, a prodrug of *nor-N*-mustard, which was therapeutically active *in vivo* (5, 8). Table 1 shows the results of early comparative experiments on Yoshida ascites sarcoma of the rat receiving a single i.v. administration of cyclophosphamide. It is evident from Table 1 that the curative activity of cyclophosphamide is about as high as that of the reference substance chlormethine *N*-oxide, but it is far less toxic than this highly reactive nitrogen mustard derivative, the result being a distinctly larger therapeutic index.

The first publications on the pharmacology of cyclophosph-



| Oxazaphosphorine               | R <sub>1</sub>   | R <sub>2</sub>                        | R <sub>3</sub>   |
|--------------------------------|--|---------------------------------------|--|
| Cyclophosphamide               | Cl-CH <sub>2</sub> -CH <sub>2</sub> -N                               | H-                                    | H-   |
| 4-Hydroperoxy-Cyclophosphamide | Cl-CH <sub>2</sub> -CH <sub>2</sub> -N                               | H-                                    | HOO-   |
| Malfosfamide                   | Cl-CH <sub>2</sub> -CH <sub>2</sub> -N                               | H-                                    | <sup>+</sup> O <sub>3</sub> S-CH <sub>2</sub> -CH <sub>2</sub> -S- |
| Ifosfamide                     | Cl-CH <sub>2</sub> -CH <sub>2</sub> -N                               | Cl-CH <sub>2</sub> -CH <sub>2</sub> - | H-   |
| Trofosfamide                   | Cl-CH <sub>2</sub> -CH <sub>2</sub> -N                               | Cl-CH <sub>2</sub> -CH <sub>2</sub> - | H-   |
| Sufosfamide                    | CH <sub>3</sub> SO <sub>2</sub> -CH <sub>2</sub> -CH <sub>2</sub> -N | Cl-CH <sub>2</sub> -CH <sub>2</sub> - | H-   |

Fig. 1. Chemical structures of particularly favorable oxazaphosphorines.

Table 1 Parameters of the curative and lethal effects of cyclophosphamide compared with chlormethine-*N*-oxide and nor-nitrogen mustard

Single i.v. administration to Yoshida's ascites sarcoma of the rat on the day of inoculation (8).

| Agent                         | LD <sub>50</sub> <sup>a</sup><br>(mg/kg) | CD <sub>50</sub><br>(mg/kg) | % of<br>LD <sub>50</sub> | Therapeutic index                    |
|-------------------------------|--|-----------------------------|--------------------------|--------------------------------------|
|                               |  |                             |                          | LD <sub>50</sub><br>CD <sub>50</sub> |
| Cyclophosphamide              | 160                                      | 4.5                         | 2.8                      | 8.5                                  |
| Chlormethine- <i>N</i> -oxide | 50                                       | 5.4                         | 10.9                     | 2.2                                  |
| nor-Nitrogen mustard          | 100                                      | 40                          | 40                       | 0.65                                 |

<sup>a</sup> LD<sub>50</sub>, 50% lethal dose; LD<sub>5</sub>, 5% lethal dose; CD<sub>50</sub>, 50% curative dose; CD<sub>95</sub>, 95% curative dose.

phamide were soon followed by confirmation of its superior chemotherapeutic properties (11, 12). The assessment of Sugiura *et al.* (13) was that "Among 1,000 selected compounds and antibiotics tested against all or portions of the tumor spectrum (33 tumors) cyclophosphamide was the most effective." The product also rapidly attracted great attention in clinical oncology. The initial clinical results from Germany (14, 15) were soon confirmed and extended internationally (16). Even now, 30 years after its introduction, cyclophosphamide is one of the most widely used cytostatics and is a constituent of many polychemotherapy regimens. There have been more than 15,000 scientific publications on this product, which demonstrate the wide interest in it. This wide interest in the unique features of cyclophosphamide not only accelerated and augmented the scientific knowledge but also yielded a number of worldwide deep personal friendships.

**Ifosfamide.** From the viewpoint of pharmacology, cyclophosphamide appeared to have reached a certain optimum status, but it subsequently emerged that ifosfamide is a cytostatic with its own remarkable chemotherapeutic properties. In ifosfamide, one of the two 2-chloroethyl groups on the extracyclic nitrogen mustard moiety in cyclophosphamide has been transferred to the nitrogen in the oxazaphosphorine ring, so that there are now two independent functional chemical groups which are further apart (Fig. 1). This apparently small difference in the chemical structure alters the metabolism *in vivo* (17) and results in a distinct change in the pharmacokinetics and pharmacodynamics (18).

With respect to practical clinical use the knowledge of the cumulative properties of both substances is of particular importance. Generally the therapeutic value of an antitumor agent is the greatest the more rapidly its toxic effect is reversible and the more its curative effect is cumulated. According to Druckrey *et al.* (11) the cumulative properties may be assessed quantitatively by determining that part which is reversible within 24 h. The cumulation residue *C* is the complement of the quantity *R* (reversible part) following the equation  $R = 1 - C$ . The *C* values can be calculated by comparative determination of the *D*<sub>50</sub> obtained with single administration *versus* a divided dose of two or more daily fractions. From this value, it is possible to calculate the height of the cumulation peak (*C*<sub>max</sub>) which is finally reached with daily administration of constant doses (11). The results of comparative studies on cyclophosphamide and ifosfamide are shown in Fig. 2. The toxic cumulation residue of ifosfamide after 24 h is about 83% and that of cyclophosphamide is nearly 100%. The curative action, assessed in Yoshida's ascitic sarcoma, behaves just the opposite; for cyclophosphamide the *C* value is about 45%, whereas for ifosfamide it is about 100%. Thus the curative action of ifosfamide is much more cumulative than that of cyclophosphamide. Accordingly on fractionated dosage, ifosfamide yielded definitively better chemotherapeutic results in tumor-bearing rats, whereas on single administration, cyclophosphamide was superior (18). This finding was the basis for the

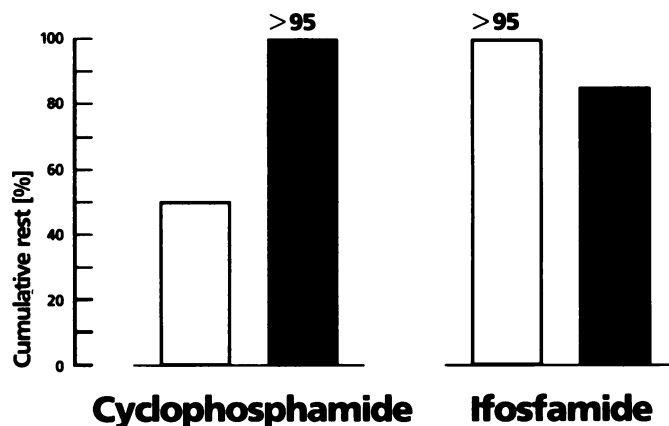


Fig. 2. Cumulative residue of the curative (□) and toxic (■) effects of cyclophosphamide and ifosfamide at 24 h after administration (2). Assay design, Sprague-Dawley rats; breeder, Mus Rattus AG, Brunnthal; sex, male; weight, 150–190 g. Standard food: Altromin 1324; water *ad libitum*, specific-pathogen-free conditions; animals per dose, 10; i.v. injection; period of observation, 28 days following the last injection; evaluation, probit method.

successful introduction of the currently used fractionated ifosfamide dose schedules in the clinic (19). Recently, this principle has been extended by administering ifosfamide as a continuous infusion for 1 to 5 days. This diminishes the toxicity, allows the total dose to be increased by nearly 50%, and improves the therapeutic results further (20, 21). The particular characteristics and clinical importance of this product, which was initially underestimated in some countries, have now received wide recognition (22–25).

#### Reliability of Experimental Findings on Cyclophosphamide and Ifosfamide to Clinical Experience

This point requires special discussion because important fundamentals of tumor pharmacology were not established until cyclophosphamide and ifosfamide were available as test substances. Both drugs were particularly suitable for a wide range of experimental and clinical uses because of their favorable physicochemical and biological properties, their high chemotherapeutic efficacy, and their relatively low toxicity. In particular, it proved possible to develop a large number of pharmacological test models for clinical oncology. These models have a crucial bridging function because they provide a scientific basis for the results of clinical trials on the effects of the drugs in patients and make them more reliable (26). They have made a considerable contribution to the development of a strategy for cancer chemotherapy.

Druckrey *et al.* (11) were aiming at optimization of the dose/time relationship when they developed massive-dose therapy. As the dose of cyclophosphamide increases, there is not only a corresponding improvement in the prospects of curing a particular tumor type but also a widening of the range of tumor types which can be influenced by the therapy. On the other hand, experiments showed that the development of secondary resistance of tumors was explained by treatment with fractionated low cyclophosphamide doses. The idea of massive-dose therapy with cyclophosphamide was taken up by clinicians. They found that the results of therapy of rapidly proliferating tumors were considerably better than with low cyclophosphamide doses (27).

Cyclophosphamide also played an important part in the development of rational polychemotherapy by Goldin *et al.* (28) and Schabel (29). When cyclophosphamide was included in new combination regimens, it was found that the clinical result depended not only on the particular chemotherapeutics chosen,

the efficacy of which varied with the type of tumor, but also on appropriate dosage and a suitable time sequence of the individual substances.

The design of scientifically based multiple chemotherapy regimens demands not only knowledge of the mechanism of action and the pharmacokinetics of the chosen cytostatics but also detailed information on the kinetics of proliferation of the tumor cells which are to be treated and of the normal cells which must be protected. There have been many studies with cyclophosphamide as the test substance which have deepened and extended our knowledge in this difficult area. The phase specificity of many cytostatics in the cell cycle has been determined, and animal models for specific sequential cytostatic treatment of tumors have been developed by Skipper (30) and his group. This has made it possible to carry out logical sequential therapy, which results in greater clinical efficacy and higher selectivity.

Cyclophosphamide and ifosfamide were also used to work out the methods for determining the cumulative properties of cytostatics (11, 31). Although cumulative properties are unwanted and hazardous when they relate to the toxic effect of chemotherapeutics, they may be beneficial when they relate to the desired therapeutic effect. Results of pharmacological studies have crucially influenced the clinical use of cyclophosphamide and ifosfamide in patients.

The fundamental studies on cyclophosphamide in postoperative and preoperative adjuvant chemotherapy are still of great importance today. Thirty years ago (32) it was shown very clearly that chemotherapy is effective as an adjuvant measure in the scheme of postoperative prophylaxis of recurrence and metastasis. Suitable experimental models have also been developed for preoperative chemotherapy (33). Thus, for example, in chemotherapeutically incurable Shay chloroma of rats, the tumor weight was greatly reduced by treatment with cyclophosphamide (30 mg/kg), and subsequent removal by surgery resulted in a high percentage of definitive cures. The importance of this animal model for clinical oncology has only recently been pointed out by Van Putten (34).

All the important organotoxic components of the effect of cyclophosphamide have been analyzed and quantitatively determined resulting in the creation of the term danger coefficient (1). This coefficient indicates the probability of particular toxic effects, *e.g.*, leukotoxicity, immunotoxicity, or vesicotoxicity associated with a particular curative dose, *e.g.*, the 95% curative dose. These data have also proved to be clinically relevant. Thus, *e.g.*, the fact that the leukotoxic effect of cyclophosphamide and ifosfamide is experimentally far less than that of directly alkylating agents has been widely confirmed in the clinic (16).

The clinical relevance of the experimental testing of cytostatics and its predictive value have been investigated by Goldin *et al.* (12); the relevance of results from animal experiments on cyclophosphamide was found to be particularly great (*i.e.*, the predictive value for the treatment of human tumors was very high). This is why cyclophosphamide was recommended by the International Conference on Screening Methodology for Antitumor Drugs as a standard against which all new developments should be compared (Geneva, 1974).

### Cytotoxic Specificity and Cancerotoxic Selectivity of Oxazaphosphorines

**Relationship to the Metabolism.** Once the relative selectivity of the oxazaphosphorine cytostatics had been demonstrated, the task was to find the biochemical and pharmacological reasons for this phenomenon and to elucidate the mechanism of action. A particularly interesting question was how a nitrogen mustard

compound is able selectively to damage tumor cells since, after all, alkylation, which is the basis for the cytotoxic action, is relatively nonspecific. Together with Hohorst, we carried out experiments attempting to explain the specificity and selectivity of alkylating nitrogen mustards, especially oxazaphosphorines (35, 36). This problem also attracted wide international interest and has been extensively investigated by many researchers, *e.g.*, Refs. 37–42.

Cyclophosphamide and other oxazaphosphorine cytostatics differ from the directly alkylating substances in that they must undergo biotransformation before they can exert their alkylating oncocidal action. The metabolism of oxazaphosphorines in warmblooded species can be divided into three stages: activation, toxification, and deactivation (36, 43) (Fig. 3). The initial activation is based on hydroxylation of the ring at C-4 to give 4-hydroxycyclophosphamide by mixed function oxidases in the liver (cytochrome P-450). The increased reactivity conferred by this enzymatic hydroxylation leads to the toxification by spontaneous elimination of acrolein and formation of the directly alkylating phosphoric acid diamide derivatives. The sequence of enzymatic activation and spontaneous, rate-limiting toxification allows intermediate reactions to take place *in vivo* on the oxazaphosphorine ring, and these result in reversible or irreversible deactivation. An irreversible enzymatic deactivation by further dehydrogenation to give the 4-keto or carboxy compound is important for excretion. This process has also been suggested as the reason for the development of resistant tumor lines (44).

A step which is crucial for the selectivity is a nonenzymatic reversible reaction of 4-hydroxycyclophosphamide with sulfhydryl compounds to give 4-thiooxazaphosphorines. Thus, for example, free sulfhydryl groups on proteins may slow down the toxification during transport in the blood and thus contribute to delaying the toxification until the tumor cell is penetrated (45).

**Cytotoxic Specificity.** When examining the problem of selectivity, the concept of cytotoxic specificity was introduced as a new and crucial parameter (36). It is defined as the biological efficacy *in vitro* of the alkylating reaction on which the cytotoxic effect depends (cytotoxic units per  $\mu\text{mol}$  and min). Surprisingly, the values for the cytotoxic specificity of the primary oxazaphosphorine metabolites are 10 to 100 times higher than those of simple alkylating agents. Comparison of these values with the therapeutic indices determined *in vivo* (corresponding to cancerotoxic selectivity) indicates (Fig. 4) that these two measures of the biological activity, which are basically independent from each other, show a largely parallel behavior. The high therapeutic indices of the oxazaphosphorines correspond to the high cytotoxic specificities of their active metabolites *in vitro*; in contrast,

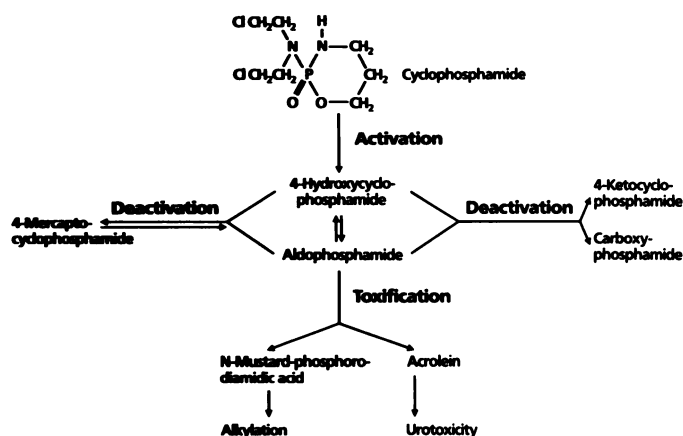


Fig. 3. Metabolic routes of activation, deactivation, and toxification of cyclophosphamide.

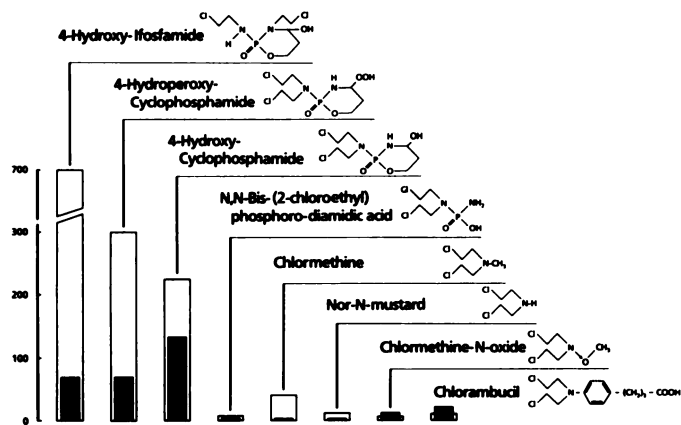


Fig. 4. Selectivity *in vivo* and specificity *in vitro* of directly alkylating *N*-mustard derivatives versus activated 2-chloroethylamino-oxazaphosphorines (36). ■, cancerotoxic selectivity (50% lethal dose: 50% curative dose); □, cytotoxic specificity, (cytotoxic units/μmol min) × 10<sup>-2</sup>.

simple nitrogen mustard derivatives have only low cancerotoxic selectivity *in vivo*, consistent with their low cytotoxic specificity. Hence the selectivity of the oxazaphosphorines *in vivo* derives mainly from the greater cytotoxic specificity of the primary metabolites and from the particular reactivity of the oxazaphosphorine ring hydroxylated on C-4.

Recent investigations by Hohorst *et al.* (45) have shown that it is possible to follow further the processes taking place in the cancer cell and thus to elucidate the actual mechanism of the high specificity and selectivity. Various 3',5'-exonucleases and phosphodiesterases catalyze the intracellular release of the alkylating metabolites from 4-hydroxycyclophosphamide, which is crucial for the oncocidal action (toxification). Particularly active 3',5'-exonucleases are those associated with DNA polymerases. They can be regarded as specific targets for the primary oxazaphosphorine metabolites. The intracellular release of the alkylating agent results in specific inhibition of the DNA polymerase enzyme moiety and thus in inhibition of DNA polymerization and to specific alkylation of the DNA.

Fig. 5 summarizes the present knowledge on the mode of action of the oxazaphosphorines and compares it with the initial idea (2). Even though the original hypothesis proved to be too simple, the high degree to which the original concept has been translated into reality is none the less remarkable. The aim of having a transport form activated enzymatically to the active form in the target organ has been achieved (as a sequence of various intermediate reactions), and the desired increase in cancerotoxic selectivity has been convincingly demonstrated. However, as the mechanism of action became clearer, and this is another benefit, new ideas for rational further development continually emerge, so that even now, 30 years after the introduction of cyclophosphamide, understanding is still increasing, and new projects are suggested; the development of stable metabolites is in progress, and there are now signs that these may be suitable for specific immunomodulation (44).

#### Organ-specific Detoxification of Urotoxic Oxazaphosphorines by Mesna

Despite all the advances, it cannot yet be said that the selectivity and therapeutic index of the oxazaphosphorines and other currently available cytostatics are satisfactory. Toxic side effects, which are often very organ specific, limit their use. Acute urotoxic effects must be expected as a consequence of the therapeutic use of the oxazaphosphorine cytostatics. These effects mainly take the form of hemorrhagic cystitis and not uncommonly limit

## IDEA

The principle of transport form (TF)/active form (AF) Nor-Nitrogen mustard

Procedure:  
Nor-Nitrogen mustard (AF)  $\xrightarrow{\text{chemical inactivation}}$  TF  
TF  $\xrightarrow{\text{enzymatical activation in vivo (tumor cells)}}$  AF

## REALITY

Nor-Nitrogen mustard  $\xrightarrow[\text{nucleophilic substitution}]{\text{chemical inactivation}}$  TF  
TF  $\xrightarrow[\text{liver microsomes}]{\text{enzymatical activation}}$  TFA (transport form of higher reactivity)  
(extracellular and intracellular (cancer))  $\text{TFA} + \text{HSR} \rightleftharpoons \text{TFA} - \text{SR}$  (temporary deactivation)  
toxification  $\downarrow$  3',5'-exonuclease / DNA-polymerase  
AF  $\xrightarrow{\text{alkylation}}$  DNA

Fig. 5. Idea and reality in the development of oxazaphosphorine cytostatics. HSR, biological thiol.

the therapy. A typical, although rare, late consequence of successful chemotherapy with cyclophosphamide which has been completed many years earlier is the occurrence of bladder carcinoma. This led to the idea of developing a uroprotector which, on systemic administration, was intended to act like an antidote by detoxifying the toxic metabolites in the kidney and urinary tract (46-49).

The urotoxicity of oxazaphosphorine cytostatics was found to be caused by the primary 4-hydroxy metabolites, specifically by the acrolein which they release. These metabolites are excreted with the urine and cause concentration-dependent damage to the kidney and bladder. An effective uroprotector ought therefore either to stabilize the 4-hydroxy metabolites in the urine, and thus prevent the release of acrolein, or to detoxify directly the acrolein being generated. A large number of substances, especially sulfur compounds, were examined for their ability to act as regional antidotes. Apart from the uroprotection, attention was directed at the pharmacokinetics, the intrinsic toxicity of the compounds, and the question of interactions. It emerged from these extensive studies that most of the tested substances were not capable of ensuring regional detoxification of the urotoxic oxazaphosphorine cytostatics. This also applies to *N*-acetylcysteine which has been repeatedly recommended for the uroprophylaxis of oxazaphosphorine cytostatics, despite clear evidence that no effective thiol concentrations appear in the urine after administration of *N*-acetylcysteine (50).

A special position is occupied by the mercaptoalkanesulfonic acids, and the compound sodium 2-mercaptoethanesulfonate (INN mesna) meets the requirements in an almost ideal manner (Fig. 6). Mesna is able to inactivate the toxic metabolites even after high doses of cyclophosphamide or ifosfamide and brings about dose-dependent prevention of the urotoxic side effects without reducing the efficacy of the cytostatic therapy itself.

The uroprotective properties of mesna are determined by its particular pharmacokinetics (51, 52). After parenteral or oral

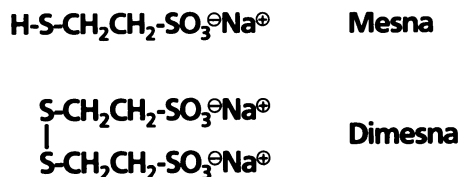


Fig. 6. Structural formulas of mesna and mesna disulfide.

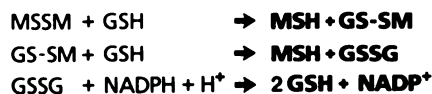


Fig. 7. Reduction of mesna disulfide into mesna by glutathione in the renal epithelia. *MSH*, mesna; *GSH*, glutathione; *MSSM*, mesna disulfide; *GSSG*, glutathione disulfide. The first two reactions are catalyzed by thioltransferase; the third reaction is catalyzed by glutathione reductase.

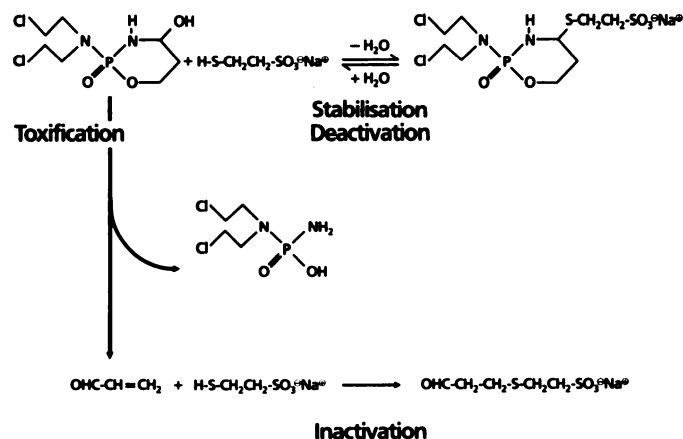


Fig. 8. Inactivation and deactivation of cyclophosphamide metabolites by mesna in the urine. Inactivation, addition reaction of mesna with the double bond of acrolein; deactivation, formation of a relatively stable and vesically nontoxic conjugate of 4-hydroxycyclophosphamide and mesna in the presence of an excess of mesna.

administration, mesna is rapidly converted in the blood into the biologically inactive mesna disulfide (dimesna), which remains in the intravascular space and is rapidly eliminated via the kidney. Following glomerular filtration, mesna disulfide enters the epithelial cells of the renal tubules where it is reduced by interaction with the glutathione system of these cells and is excreted with the urine as a reactive thiol compound (Fig. 7). Only this detoxifies the aggressive oxazaphosphorine metabolites in the urine (51) (Fig. 8).

In clinical practice the administration of mesna prevents the urotoxic side effects of oxazaphosphorine therapy allowing higher doses of these cytostatics to be given, so that the therapeutic efficacy is improved and new indications have emerged (23–25, 53).

Experimentally, it was found that mesna can also inhibit or suppress the occurrence of bladder tumors when administered simultaneously during long-term treatment with cyclophosphamide (54). It is to be expected that effective prophylaxis of this tragic late effect will also be possible clinically. Thus, in future, the therapeutic use of oxazaphosphorines, especially cyclophosphamide and ifosfamide, should always be combined with uroprotection in order to circumvent reliably the risk of inflammation and of urinary bladder tumors.

#### Future Prospects in Oxazaphosphorine Research

The metabolites of the oxazaphosphorine cytostatics which play a key part are the 4-hydroxy compounds. They are the only ones which share the cytotoxic specificity *in vitro* and the can-

cerotoxic selectivity *in vivo* with the parent substances; in this respect they differ very considerably from all the directly alkylating compounds. The difference from the parent substances, e.g., cyclophosphamide, is that they no longer require enzymatic activation for their action; the release of the primary activated metabolite (e.g., 4-hydroxycyclophosphamide) takes place spontaneously in aqueous solution and depends on the chemical reactivity of the particular compound and the conditions prevailing, such as the pH and temperature. The pharmacokinetics and the biological variability in both the initial enzymatic activation and the competing deactivation reactions made it appear desirable to test the 4-hydroxy metabolites therapeutically. For a long time this was impossible due to the chemical instability. It has recently been possible to synthesize a number of new and more stable compounds by replacement of the 4-hydroxy group, and these are sufficiently stable in the crystalline form at room temperature but spontaneously liberate the 4-hydroxy compound in aqueous solution (5).

The prototype of this class of substances is mafosfamide (Asta Z 7557, Z 7654) which has 2-mercaptoethanesulfonate as substituent in position 4 of the oxazaphosphorine ring (Fig. 1). The drug offers new and unexpected therapeutic approaches (44). Pretherapeutic evaluation in clonogenic stem cell assays had been impossible with the parent compounds, which required enzymatic activation. Intracavitary administration or site-directed perfusion can be carried out with mafosfamide. An interesting aspect now under clinical investigation is the extracorporeal purging of bone marrow before autologous transplantation in acute leukemia. Even more important in the near future might be the evaluation of the immunomodulating potency of low-dose mafosfamide, which in some experimental models induces permanent cures from cancer, simply by modulation of the host's defense system. An increase of this type in the immune response has also been reported with low cyclophosphamide or ifosfamide doses. Analysis of these findings by immunological methods suggested that the therapeutic efficacy of oxazaphosphorines in low, nontoxic doses derived from elimination of suppressor mechanisms (55). Mafosfamide, the stable metabolite, is very suitable for preclinical and clinical immunopharmacological research. The first clinical investigations of this new and fascinating aspect of oxazaphosphorine cytostatics have been initiated. These will also examine whether and how this new principle can be utilized therapeutically in conjunction with "biological response modifiers" (44, 56).

#### Conclusion

The research results which I have summarized arose from the desire to increase the selectivity of cancer chemotherapy. I would like to conclude with the forward-looking thoughts of Dr. Gertrude B. Elion, a past recipient of the Bruce F. Cain Award: "Chemotherapeutic agents are not only ends in themselves, they are also beginnings. . . . Selectivity must be our goal and understanding its basis our guide for the future." Further advances will depend more than ever on our intuition and the courage of new ideas, since, as Elion says "the basic knowledge and serendipity are not opposites but integral parts of the process of discovery" (57).

#### References

1. Brock, N. Experimental basis of cancer chemotherapy. In: K. Hellmann, and T. A. Connors (eds.), *Chemotherapy*, Vol. 7. New York: Plenum Publishing Corp., 1976.
2. Brock, N. Ideas and reality in the development of cancer chemotherapeutic



- agents, with particular reference to oxazaphosphorine cytostatics. *J. Cancer Res. Clin. Oncol.*, **111**: 1-12, 1986.
3. Heubner, W. Einfall und Zufall bei der Auffindung von Arzneimitteln. *Pharmazie*, **101**: 378, 1956.
4. Druckrey, H. Experimentelle Grundlagen der Chemotherapie des Krebses. *Dtsch. Med. Wochenschr.*, **77**: 1534-1537, 1952.
5. Brock, N. Oxazaphosphorine cytostatics: structure-activity relationships, selectivity and metabolism, regional detoxification. In: D. N. Reinholdt, T. A. Connors, H. M. Pinedo, and K. W. Van de Poll (eds.), *Structure-Activity Relationships of Anti-Tumour Agents*, pp. 239-267. The Hague: Martinus Nijhoff, 1983.
6. Ishidate, M., Sakurai, Y., Yoshida, T., Sato, H., Sato, H., and Matsui, E. Experimental studies on chemotherapy of malignant growth employing Yoshida sarcoma animals. *Gann*, **44**: 342-346, 1953.
7. Friedman, O. M., and Seligman, A. M. Preparation of *N*-phosphorylated derivatives of bis- $\beta$ -chloroethylamine. *J. Am. Chem. Soc.*, **76**: 655-658, 1954.
8. Brock, N. Zur pharmakologischen Charakterisierung zyklischer *N*-Lost-Phosphamidester als Krebs-Chemotherapeutika. *Arzneim.-Forsch./Drug Res.*, **8**: 1-9, 1958.
9. Arnold, H., and Bourseaux, F. Synthese und Abbau zytostatisch wirksamer zyklischer *N*-Phosphamidester des Bis-( $\beta$ -chloräthyl)-amins. *Angew. Chem.*, **70**: 539-544, 1958.
10. Arnold, H., Bourseaux, F., and Brock, N. Über Beziehungen zwischen chemischer Konstitution und cancerotoxischer Wirkung in der Reihe der Phosphamidester des Bis-( $\beta$ -chloräthyl)-amins. *Arzneimittelforsch./Drug Res.*, **11**: 143-158, 1961.
11. Druckrey, H., Steinhoff, F., Nakayama, M., Preussmann, R., and Anger, K. Experimentelle Beiträge zum Dosisproblem in der Krebs-Chemotherapie und zur Wirkungsweise von Endoxan. *Dtsch. Med. Wochenschr.*, **88**: 651, 1963.
12. Goldin, A., Serpick, A. A., and Mantel, N. Experimental screening procedures and clinical predictability value. *Cancer Chemother. Rep.*, **50**: 173-218, 1966.
13. Sugiura, K., Schmid, F. A., and Schmid, M. M. Antitumor activity of cytoxan. *Cancer Res.*, **21**: 1412-1420, 1961.
14. Wilmanns, H. *Asta-Forschung und Therapie*, 1958.
15. Gross, R., and Wulf, G. Klinische und experimentelle Erfahrungen mit zyklischen und nichtzyklischen Phosphamidestern des *N*-Lost in der Chemotherapie von Tumoren. *Strahlentherapie*, **41** (Sonderband III): 361-367, 1959.
16. Hill, D. L. *A Review of Cyclophosphamide*. Springfield, IL: Charles C Thomas, Publisher, 1975.
17. Norpoth, K. A Contribution to the metabolism of ifosfamide in the rat and man. In: *Proceedings of the International Holoxan Symposium*, Düsseldorf, Germany, 1977, pp. 40-45.
18. Brock, N. Pharmacological studies with ifosfamide, a new oxazaphosphorine compound. In: M. Semonsky, M. Hijzlar, and S. Masak (eds.), *Advances in Antimicrobial and Antineoplastic Chemotherapy*, pp. 759-761. Munich; Urban & Schwarzenberg, 1972.
19. Schnitker, J., Brock, N., Burkert, H., and Fichtner, E. Evaluation of a cooperative clinical study of the cytostatic agent ifosfamide. *Arzneimittelforsch./Drug Res.*, **26**: 1783-1793, 1976.
20. Klein, H. O., Christian, E., Coerper, C., and Wickramanayake, P. D. High-dose ifosfamide (IFO) and mesna as continuous infusion over five days—a phase I/II trial. In: *Thirteenth International Cancer Congress*, Seattle, 1982.
21. Wiltshaw, E., and Stuart-Harris, R. Single agent ifosfamide plus mesna for metastatic soft tissue sarcoma. In: *Thirteenth International Cancer Congress*, Seattle, 1982.
22. Spitz, K. H., and Karrer, K. Oxazaphosphorine—safety and therapeutic potential. In: *Proceedings of the Thirteenth International Congress of Chemotherapy*, Vienna, Symposium 83, 1983.
23. Van Oosterom, A. T., and Einhorn, L. H. Recent experience with ifosfamide—mesna in solid tumors. In: *Third European Conference on Clinical Oncology and Cancer Nursing. Satellite Symposium*, Stockholm, June 17, 1985.
24. Brade, W. P., Herdrich, K., and Varini, M. Ifosfamide—pharmacology, safety and therapeutic potential. *Cancer Treat. Rev.*, **12**: 1-47, 1985.
25. Brade, W. P., Nagel, G. A., and Seeber, S. Ifosfamide in tumor therapy. *Beitr. Onkol.*, **26**: 1-455, 1987.
26. Brock, N., and Schneider, B. Modelle in der zytostatischen Chemotherapie. In: R. Gross (ed.), *Modelle und Realitäten in der Medizin*, pp. 127-146. Stuttgart: F. K. Schattauer Verlag, 1983.
27. Schmitz, G., and Gross, R. Erste klinische Erfahrungen mit ultrahohen Einzeldosen von Cyclophosphamid. *Med. Welt*, **18**: 985-989, 1967.
28. Goldin, A., Venditti, J. M., and Mantel, N. Combination Chemotherapy: basic considerations. In: Heffter-Heubner (eds.), *Handbuch der experimentellen Pharmakologie*, Vol. 38/1, p. 411. Berlin: Springer-Verlag, 1974.
29. Schabel, F. M., Jr. New experimental drug combinations with potential clinical utility. *Biochem. Pharmacol.*, **23** (Suppl. 2): 163-176, 1974.
30. Skipper, H. E. The cell cycle and chemotherapy of cancer. In: R. Baserga (ed.), *The Cell Cycle and Cancer*, p. 355. New York: Marcel Dekker, 1971.
31. Brock, N., and Schneider, B. Models and methods for the assessment of cumulation of drug effects. *Arzneim.-Forsch./Drug Res.*, **30**: 1034-1040, 1980.
32. Druckrey, H., Kuk, B. T., Schmäh, D., and Steinhoff, D. Kombination von Operation und Chemotherapie beim Krebs. *Münch. Med. Wochenschr.*, **100**: 1913-1918, 1958.
33. Brock, N. Neue experimentelle Ergebnisse mit *N*-Lost-Phosphamidestern. *Strahlentherapie*, **41**: 347-354, 1959.
34. Van Putten, L. M. Experimental preoperative chemotherapy. *Recent Results Cancer Res.*, **103**: 36-50, 1986.
35. Hohorst, H. J., Draeger, U., Peter, G., and Voelcker, G. The problem of oncostatic specificity of cyclophosphamide (NSC-26271): studies on reactions that control the alkylating and cytotoxic activity. *Cancer Treat. Rep.*, **60**: 309-315, 1976.
36. Brock, N., and Hohorst, H. J. The problem of specificity and selectivity of alkylating cytostatics: studies on *N*-2-chloroethylamido-oxazaphosphorines. *Z. Krebsforsch.*, **88**: 185-215, 1977.
37. Struck, R. F., Kirk, M. C., and Mellett, L. B. Urinary metabolites of the antitumor agent cyclophosphamide. *Mol. Pharmacol.*, **7**: 519-529, 1971.
38. Sladek, N. E. Bioassay and relative cytotoxic potency of cyclophosphamide metabolites generated *in vitro* and *in vivo*. *Cancer Res.*, **33**: 1150-1158, 1973.
39. Connors, T. A., Cox, P. J., Farmer, P. B., Foster, A. B., and Jarman, M. Some studies of the active intermediates formed in the microsomal metabolism of cyclophosphamide and isophosphamide. *Biochem. Pharmacol.*, **23**: 115-129, 1974.
40. Foster, A. B., Farmer, P. B., Connors, T. A., Cox, P. J., and Jarman, M. Symposium on the Metabolism and Mechanism of Action of Cyclophosphamide, Chester Beatty Research Institute, London, England, 1975, published in *Cancer Treat. Rep.*, **60**: 299-525, 1976.
41. Colvin, M. The comparative pharmacology of cyclophosphamide and ifosfamide. *Semin. Oncol.*, **9**: 2-7, 1982.
42. Kohn, F. R., and Sladek, N. E. Aldehyde dehydrogenase activity as the basis for the differential sensitivity of murine pluripotent and progenitor hematopoietic cells to oxazaphosphorine *in vitro*. *Proc. Am. Assoc. Cancer Res.*, **25**: 289, 1984.
43. Brock, N., and Hohorst, H. J. Über die Aktivierung von Cyclophosphamid im Warmblüterorganismus. *Naturwissenschaften*, **49**: 610-611, 1962.
44. Brock, N., Hilgard, P., Peukert, M., Pohl, J., and Sindermann, H. Basis and new developments in the field of oxazaphosphorines. *Cancer Invest.*, in press, 1988.
45. Hohorst, H. J., Bielicki, L., and Voelcker, G. The enzymatic basis of cyclophosphamide specificity. *Adv. Enzyme Regul.*, **25**: 99-122, 1986.
46. Brock, N. Konzeption und Wirkmechanismus von Uromitexan (Mesna). *Beitr. Onkol.*, **5**: 1-11, 1980.
47. Brock, N., Pohl, J., and Stekar, J. Studies on the urotoxicity of oxazaphosphorine cytostatics and its prevention. I. Experimental studies on the urotoxicity of alkylating compounds. *Eur. J. Cancer*, **17**: 595-607, 1981.
48. Brock, N., Pohl, J., and Stekar, J. Studies on the urotoxicity of oxazaphosphorine cytostatics and its prevention. II. Comparative study on the uroprotective efficacy of thiols and other sulfur compounds. *Eur. J. Cancer Clin. Oncol.*, **17**: 1155-1168, 1981.
49. Brock, N., Pohl, J., and Stekar, J. Studies on the urotoxicity of oxazaphosphorine cytostatics and its prevention. III. Profile of action of sodium 2-mercaptoethane sulfonate (mesna). *Eur. J. Cancer Clin. Oncol.*, **18**: 1377-1387, 1982.
50. Brock, N., and Pohl, J. Organspezifische Entgiftung von Zytostatika. Wirkungsvergleich Mesna—*N*-Acetylcystein. *Therapiewoche*, **37**: 2403-2407, 1987.
51. Ormstad, K., Orrenius, S., Lastbom, T., Uehara, N., Pohl, J., Stekar, J., and Brock, N. Pharmacokinetics and metabolism of sodium 2-mercaptoethane-sulfonate in the rat. *Cancer Res.*, **43**: 333-338, 1983.
52. Brock, N., Hilgard, P., Pohl, J., Ormstadt, K., and Orrenius, S. Pharmacokinetics and mechanism of action of detoxifying low-molecular-weight thiols. *J. Cancer Res. Clin. Oncol.*, **108**: 87-97, 1984.
53. Burkert, H., and Nagel, G. A. (eds.). *New Experience with Oxazaphosphorines with Special Reference to the Uroprotector Uromitexan®*. Basel: S. Karger AG, 1981.
54. Brock, N., Habs, M., Pohl, J., Schmäh, D., and Stekar, J. Mesna (Natrium-2-Mercaptoethansulfonat). Prophylaxe der akuten urotoxischen Nebenwirkungen sowie der kanzerogenen Spätfolgen von Cyclophosphamid und Ifosfamid. *Therapiewoche*, **32**: 4975-4996, 1982.
55. Ozer, H., Cowens, J. W., Colvin, M., Nussbaum-Blumenson, A., and Sheedy, D. *In vitro* effects of 4-hydroperoxycyclophosphamide on human immunoregulatory T subset function. I. Selective effects on lymphocyte function in T-B collaboration. *J. Exp. Med.*, **155**: 276-290, 1982.
56. Hilgard, P., Pohl, J., Stekar, J., and Voegeli, R. Oxazaphosphorines as biological response modifiers—experimental and clinical perspectives. *Cancer Treat. Rev.*, **12**: 155-162, 1985.
57. Elion, G. B. Selectivity—key to chemotherapy: Presidential Address. *Cancer Res.*, **45**: 2943-2950, 1985.